

***KRISHNA VISHWA VIDYAPEETH (DEEMED TO BE UNIVERSITY), KARAD***

***Accredited By NAAC With 'A+' Grade***



***Revised Syllabus (CBCS) For***  
***Bachelor of Science Food and Dairy***

## **Prologue**

The Institute of Allied Sciences was established in 2007 with Five Under graduate courses Microbiology, Food and Dairy, Environmental Science, Nutrition & Dietetics and Food & Dairy Technology. Currently Eighteen faculty members are engaged in Academic functions.

The seemingly overwhelming and ever-expanding state of knowledge about microorganisms, their genetic material, Molecular Biology and Recombinant DNA Technology increases the scope of Food and Dairy. This newly emerging branch of science offers something for everyone and it cultivates informed citizens who can make perceptive decisions on important events. Many discoveries made by Microbiologists and Biotechnologists have spawned new fields of science such as molecular Biology, Genetics, Enzyme Technology, Fermentation Technology, Bioengineering, Genetic Engineering, Immunology etc. Many studies have been made using Science and Food and Dairy to understand the principles that govern life.

New developments are occurring constantly in these areas and thus Biotechnologies have become the mainstays of many technologies. This has necessitated the formation of the Food and Dairy courses for the development of competent, smart and dynamic Biotechnologists that are required in Academic Institutes, Research organizations, Professional organizations and in various industries such as Pharmaceutical Industries, Enzyme Industries, Food and Dairy Industries, Wine and Alcohol Industries, Agro based Industries. **The Choice –Based Credits System(CBCS)** provides for a framework within which there is flexibility in the design of courses and their content, simultaneously also providing the students a choice of the courses he/she wishes to study. The courses are assigned credits based on teaching hours, which in turn is linked to courses content and structure

The rapid pace of discovery and their application dictates a somewhat selective inclusion of theory paper / topics and practical and proper training of the students. The course is designed in such a way that students remain constantly busy with their studies through the Lecture and Practical periods, Seminar periods, Home assignments, Mid – term examinations (Periodic tests), Preliminary or term end examinations and also gets exposure to outside world through visits to Research Laboratories / Science Institutes / Industries of Microbiological /Biotechnological interest. The course also makes the provision for training in research through the research project (during one or two semesters) and / or Industrial training in organization of Microbiological interest. (During one semester / one summer vacation.)

Over all it is aimed to design **Three year under graduate (B.Sc.) course in Food and Dairy** with a balanced coverage of traditional and “cutting edge technology” along with the necessary courses (Communication skills, Biostatistics, Computer science, Scientific writing and Presentation, Research training / Industrial training) as per the UGC guidelines and produce competent Biotechnologists to meet the demand of Industries, Research organizations and Academic Institutes in the country and abroad.

### **Process of Curriculum Design**

**The Choice-Based Credit System (CBCS)** provides a framework within which there is flexibility in the design of courses and their content. At the same time it also provides the student a choice of the courses he/she wishes to study. The courses are assigned credits based on teaching hours, which in turn is linked to course content and structure.

### **Curriculum Designing Process**

Following procedure was adopted for curriculum designing: For curriculum development first need analysis was done and then based on need analysis draft syllabus was prepared in the Departmental Curriculum Committee meeting and it was subsequently discussed in College Curriculum Committee meeting where all faculty members participated in the discussion and debated over the draft syllabus. The draft syllabus approved in the College Curriculum Committee meeting was sent to BOS where inputs given by external subject experts were considered and incorporated in the final draft. The draft syllabus finalized in BOS was sent to Academic Council for its approval.

When revising the syllabi for the courses, the courses to be implemented as well as the content of each course was extensively discussed and debated on, feedback obtained from students, faculty, subject experts from academic institutes, industry experts, alumni were extensively discussed and debated in the meetings of curriculum committees and BOS and the inputs were considered. Thus for the development of syllabus contributions came from external subject experts, faculty members, feedback obtained from students, alumni, external experts and members of industry.

### **B.Sc. Food and Dairy program objectives**

After completion, the students are expected to understand the:

- a) Basic and applied aspects of microbial diversity and systematic taxonomy, Physiology, biochemistry and applications of basic aspects of microbial diversity.
- b) Principles, working and application of bioinstruments used in isolation and identification of microbes and structural determination of biomolecules,
- c) characteristics and significance of archaea, algae, fungi, viruses,
- d) Impact of various groups of microbes on earth atmosphere, human, plant and animal health and technology development,
- e) structure, properties, pathways, significance and applications of microbial biomolecules,
- f) basic and applied aspects of Genetic makeup of bacteria, algae, fungi and viruses,
- g) causes, mechanisms and consequences of defect in gene/genome of microorganisms, and
- h) basic concepts of microbial enzymes, enzyme kinetics, regulation of enzyme activity, industrial applications of enzymes, enzyme function in non-aqueous environment.

**Structure of B.Sc. program in Food and Dairy** B.Sc. **Food and Dairy** program is of two years duration and is conducted in four semesters. As recommended by UGC university has adopted a outcome-based education approach. The various courses of the program are designed to include classroom teaching, laboratory work, project work, seminars, home assignments, industrial visit etc.

### **Program Educational Objectives:**

The objectives of the **B. Sc. Programme in Food and Dairy** is:

- i) To equip the students with the basic and applied knowledge of molecular mechanisms of cellular processes in living systems including microbes, plants, animals and humans.
- ii) To provide the students with laboratory (experimental ) training so that they are competent enough to work in industries.
- iii) To provide the students with the current updates in the areas of Analytical Techniques , Industrial Fermentations, Environmental Food and Dairy.
- iv) To train students with research work methodology through small project work.
- v) To generate competent skilled human resource for industries and research organization.

### **Eligibility**

Candidates must have passed B.Sc. With minimum 50% marks with Food and Dairy/ Microbiology/ Industrial Microbiology/ Zoology/Botany as principal subject or with Biochemistry/ Microbiology/ Botany/ Zoology as subsidiary subjects at B.Sc. II level

### **Course fees**

As shown in Admission Broacher of respective year (Subject to change as and when required)

### **Duration**

The duration of B.Sc. (Microbiology) degree program shall consist of two academic years divided in to four semesters. Each Semester consist of 90 working days. Each theory and practical course must be completed in 60 lectures/Practical periods, respectively of 60 min duration.

### **Medium of instruction**

The medium of instruction and examination for each course shall be English.

### **Credit to contact hour**

One credit is equivalent to 15 periods of 60 minutes each for theory course lecture. While credit weightage for self-learning based on e-content shall be 50% or less than that for lectures.

### **Attendance**

The student enrolled for B.Sc. Food and Dairy must have 75% attendance in each course in order to appear for term end examinations, otherwise the candidate may not be allowed to appear for term end examination as per ordinance.

1] The entire B.Sc. course in Food and Dairy shall be covered in 28 [Twenty Eight] theory papers, 28 [Twenty Eight] practical courses [semester I, II, III] and a project work / Industrial training [in lieu of one practical courses of semester IV] each semester there shall be four theory papers each carrying 100 marks and for first three semesters viz. semester I, II and III, there shall be two practical courses each practical course shall carry 100 marks. However, for semester IV there shall be a research project work / Industrial training of 100 [one hundred] marks in lieu of one practical course in addition to four-theory paper and one practical course.

Semester I: Eight theory papers and Eight practical courses.

Semester II: Eight theory papers and Eight practical courses.

Semester III: Six theory papers and Six practical courses.

Semester IV: Six theory papers. Six practical course and a project work/Industrial training practical course for every student.

2] Each theory paper will be covered in four lectures of 60 minutes each per week.

Practical course shall be covered in 04 practical turns of 04 clock hours practical periods per week.

3] A practical batch shall be of 12 [twelve] to 15 [fifteen] students.

4] For university practical examination the duration should be as shown below,

For every semester there shall be two/three days practical examination for not less than 5 ½ hours.

5] Each candidate must produce a certificate from the Head of the Department in his/her college / Institute / University stating that he/she has completed, in a satisfactory manner, a practical course on the lines laid down from time to time by Academic Council on the recommendations of Board of studies and that the laboratory journal has been properly maintained. Every candidate must have recorded his/her observation in the laboratory journal and a written report on each exercise performed. Every journal is to be checked and signed periodically by a member of teaching staff and certified by the Head of the Department at the end of each semester. Candidates are to produce their journal at the time of practical examination.

- 6] There shall be one compulsory seminar of minimum 15 min. delivery per paper per semester for each student and there shall be two marks for each seminar in Internal evaluation.

During semester I & II students shall have to undertake an academic tour to visit a minimum one place of academic interests like Academic Institute/ Research Institution / R&D Department/Industry. The student should submit the report of their visit at the time of practical examination. The report should be duly certified by the Head of the Department of Microbiology, Food and Dairy.

- 7] During semester Student is to undertake a research project [as part of the semester IV] which is to be started in the beginning of semester III so as to give enough time for duly completion of project. In the project student is to study research methodology Information collection (reference work) selection of topic, outline of the work, thinking and planning, project report writing in the form of dissertation or small Project Report and the submission of the project report [Introduction, Aims and objectives, Material and method, Results and Discussions, summary, Conclusions and Bibliography] For the research project work out of one hundred marks, fifty marks shall be given by university examiners though assessment of Project Report at the time of semester IV practical examination. The remaining fifty marks shall be given by the Committee for Internal Evaluation of Projects (CIEP) as an internal evaluation. CIEP is to be constituted by the Principal (and which shall be consisting of HOD, Guide/Teacher in - charge and at least one other faculty members). The method and process of Internal evaluation is to be worked out by the CIEP.
- a) \*\*The Institute or guide of student should locate the industry and depute the student in the industry for the period of one month.
  - b) Student should complete its industrial training cum industrial project in the vacation period after semester II
  - c) Student should study microbiological and / or biotechnological aspects in industry and submit its report in the form of dissertation or small Project Report duly signed by industry authority, concerned guide and Head of the Department of Microbiology, Food and Dairy.

**Three-year**  
**B.Sc. Food and Dairy Technology**  
**Programme**  
**Course Structure**

**B.Sc. Food and Dairy Technology Part I, Semester I**

<b>B.Sc. Food and Dairy Technology Part I, Semester I (w.e.f. 2022-2023)</b>													
S r · N o	Course Code	Course Title	Teaching Hours/ Week			Marks				Credits			
			T	P	To tal	Inter nal		Exter nal			To tal		
						T	P	T	P				
<b>CGPA Theory Courses</b>													
<b>CG PA</b>	1	UG FD – T101 CC	Fundamentals of Microbial and Biological World		2	-	2	1 0	-	4 0	-	50	2
	2	UG FD – T102 CC	Fundamentals of Physics and Biophysics for Biologists		2	-	2	1 0	-	4 0	-	50	2
	3	UG FD – T103 CC	Fundamentals of Chemistry for Biologists		2	-	2	1 0	-	4 0	-	50	2
	4	UG FD – T104 CC	Fundamentals of Biosciences – Botany and Zoology		2	-	2	1 0	-	4 0	-	50	2
	5	UG FD – T105 CC	Basics of Bacteriology, Virology and Rickettsiology		2	-	2	1 0	-	4 0	-	50	2
	6	UG FD – T106 CC	Basics of Mycology, Phycology and Protozoology		2	-	2	1 0	-	4 0	-	50	2
	7	UG FD – T107 CCS	Introduction to the world of amazing microorganisms		2	-	2	1 0	-	4 0	-	50	2
	8	UG FD – T108 DSC	Basics techniques in Microbiology, Food and Dairy and Environmental Sciences		2	-	2	1 0	-	4 0	-	50	2
<b>CGPA Practical Courses</b>													
<b>CG PA</b>	9	UG FD – P101 CC	Practicals related to the theory paper - Fundamentals of Microbial and Biological World		-	2	2	-	1 0	-	40	50	1
	10	UG FD – P102 CC	Practicals related to the theory paper - Fundamentals of Physics and Biophysics for Biologists		-	2	2	-	1 0	-	40	50	1
	11	UG FD – P103 CC	Practicals related to the theory paper - Fundamentals of Chemistry for Biologists		-	2	2	-	1 0	-	40	50	1
	12	UG FD – P104 CC	Practicals related to the theory paper - Fundamentals of Biosciences – Botany and Zoology		-	2	2	-	1 0	-	40	50	1
	1	UG FD –	Practicals related to the		-	2	2	-	1	-	40	50	1



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	3	P105 CC	theory paper - Basics of Bacteriology, Virology and Rickettsialogy					0				
	1 4	UG FD – P106 CC	Practicals related to the theory paper - Basics of Mycology, Phycology and Protozoology	-	2	2	-	1 0	-	40	50	1
	1 5	UG FD – P107 CCS	Practicals related to the theory paper - Introduction to the world of amazing microorganisms	-	2	2	-	1 0	-	40	50	1
	1 6	UG FD – P108 DSC	Practicals related to the theory paper - Basics techniques in Microbiology, Food and Dairy and Environmental Sciences	-	2	2	-	1 0	-	40	50	1
	<b>Total</b>			16	16	32	8 0	8 5	3 2 0	32 0	80 0	24
<b>Mandatory Non CGPA Courses</b>												
<b>Non - CG PA</b>	1 8	UG FD – T109 SECC	Yoga and Meditation	0.5	-	0.5	2 5	-	-	-	25	0.5
	1 9	UG FD – T110 AECC	Spoken English - I	0.5	-	0.5	2 5	-	-	-	25	0.5
	<b>Total</b>			1	-	1	5 0	-	-	-	50	1
<b>Total Credits for Semester I : 25 (T = Theory: 16, P = Practical : 8, Non-CGPA : 1)</b> <b>CC : Core Course, CCS : Core Course Specialization, DSC : Discipline Specific Course,</b> <b>DSE : Discipline Specific Elective,</b> <b>SECC = Skill Enhancement Compulsory Course : 0.5, AECC = Ability Enhancement</b> <b>Compulsory Course : 0.5,</b> <b>Total Credits for Semester I CGPA Course = 24 credits</b>												

**B.Sc. Food and Dairy Technology Part I, Semester II**

<b>B.Sc. Food and Dairy Technology Part I, Semester II (w.e.f. 2022-2023)</b>												
S r · N o	Course Code	Course Title	Teaching Hours/ Week			Marks				Cre dits		
			T	P	To tal	Inter nal		Exter nal			To tal	
						T	P	T	P			
<b>CGPA Theory Courses</b>												
<b>CG PA</b>	1	UG FD – T201 CC	Basics of Cell Biology and Physiology	2	-	2	1 0	-	4 0	-	50	1.5
	2	UG FD – T202 CC	Basics of Biochemistry – Biomolecules - I	2	-	2	1 0	-	4 0	-	50	1.5
	3	UG FD – T203 CC	Basics of Biochemistry – Biomolecules - II	2	-	2	1 0	-	4 0	-	50	1.5
	4	UG FD – T204 CC	Microbial Nutrition and Growth	2	-	2	1 0	-	4 0	-	50	1.5
	5	UG FD – T205 CC	Advanced Chemistry and Physics for Biologists	2	-	2	1 0	-	4 0	-	50	1.5
	6	UG FD – T206 CC	Applied Plant and Animal Sciences	2	-	2	1 0	-	4 0	-	50	1.5
	7	UG FD – T207 CCS	Basics of Ecology, Ecosystem and Geosciences	2	-	2	1 0	-	4 0	-	50	1.5
	8	UG FD – T208 DSC	Applied Microbiology and Basics of Environmental Pollution	2	-	2	1 0	-	4 0	-	50	1.5
<b>CGPA Practical Courses</b>												
<b>CG PA</b>	9	UG FD – P201 CC	Practicals related to the theory paper - Basics of Cell Biology and Physiology	-	2	2	-	1 0	-	40	50	1
	10	UG FD – P202 CC	Practicals related to the theory paper - Basics of Biochemistry – Biomolecules - I	-	2	2	-	1 0	-	40	50	1
	11	UG FD – P203 CC	Practicals related to the theory paper - Basics of Biochemistry – Biomolecules - II	-	2	2	-	1 0	-	40	50	1
	12	UG FD – P204 CC	Practicals related to the theory paper - Microbial Nutrition and Growth	-	2	2	-	1 0	-	40	50	1
	1	UG FD –	Practicals related to the	-	2	2	-	1	-	40	50	1

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	3	P205 CC	theory paper - Advanced Chemistry and Physics for Biologists					0				
	14	UG FD – P206 CC	Practicals related to the theory paper - Applied Plant and Animal Sciences	-	2	2	-	10	-	40	50	1
	15	UG FD – P207 CCS	Practicals related to the theory paper - Basics of Ecology, Ecosystem and Geosciences	-	2	2	-	10	-	40	50	1
	16	UG FD – P208 DSC	Practicals related to the theory paper - Applied Microbiology and Basics of Environmental Pollution	-	2	2	-	10	-	40	50	1
<b>CGPA</b>	17	UG FD – P209 PP	Project I	-	1	1	-	5	-	-	5	1
			<b>Total</b>	16	17	33	80	85	320	320	805	21
<b>Mandatory Non CGPA Courses</b>												
<b>Non - CGPA</b>	18	UG FD – T209 SECC	Soft Skill and Personality Development	0.5	-	0.5	25	-	-	-	25	0.5
	19	UG FD – T210 AECC	Spoken English – II (Communication Skills)	0.5	-	0.5	25	-	-	-	25	0.5
			<b>Total</b>	1	-	1	50	-	-	-	50	1
<p style="text-align: center;"><b>Total Credits for Semester II : 22 (T = Theory: 12, P = Practical : 8, Project : 1, Non-CGPA : 1)</b></p> <p style="text-align: center;"><b>CC : Core Course, CCS : Core Course Specialization, DSC : Discipline Specific Course, DSE : Discipline Specific Elective, PP : Project</b></p> <p style="text-align: center;"><b>SECC = Skill Enhancement Compulsory Course : 0.5, AECC = Ability Enhancement Compulsory Course : 0.5,</b></p> <p style="text-align: center;"><b>Total Credits for Semester II CGPA Course = 21 credits</b></p>												

**B.Sc. Food and Dairy Technology Part II, Semester III**

<b>B.Sc. Food and Dairy Technology Part II, Semester III (w.e.f. 2023-2024)</b>												
S r . N o	Course Code	Course Title	Teaching Hours/ Week			Marks				Cre dits		
			T	P	To tal	Inter nal		Exter nal			To tal	
						T	P	T	P			
<b>CGPA Theory Courses</b>												
<b>CG PA</b>	1	UG FD – T301 CC	Food Engineering	2	-	2	1 0	-	4 0	-	50	2.5
	2	UG FD – T302 CC	Dairy Engineering	2	-	2	1 0	-	4 0	-	50	2.5
	3	UG FD – T303 CC	Food Process Technology	2	-	2	1 0	-	4 0	-	50	2.5
	4	UG FD – T304 CC	Food and Dairy Process Technology – I (Utility)	2	-	2	1 0	-	4 0	-	50	2.5
	5	UG FD – T305 CCS	Food and Dairy Process Technology – II (Equipments)	2	-	2	1 0	-	4 0	-	50	2.5
	6	UG FD – T306 DSC	Mathematics, Statistics and Computer Applications in Food and Dairy Technology	2	-	2	1 0	-	4 0	-	50	2.5
<b>CGPA Practical Courses</b>												
<b>CG PA</b>	7	UG FD – P301 CC	Practicals related to the theory paper - Food Engineering	-	1	1	-	1 0	-	40	50	1
	8	UG FD – P302 CC	Practicals related to the theory paper - Dairy Engineering	-	1	1	-	1 0	-	40	50	1
	9	UG FD – P303 CC	Practicals related to the theory paper - Food Process Technology	-	1	1	-	1 0	-	40	50	1
	10	UG FD – P304 CC	Practicals related to the theory paper - Food and Dairy Process Technology – I (Utility)	-	1	1	-	1 0	-	40	50	1
	11	UG FD – P305 CCS	Practicals related to the theory paper - Food and Dairy Process Technology – II (Equipments)	-	1	1	-	1 0	-	40	50	1
	12	UG FD – P306 DSC	Practicals related to the theory paper - Mathematics, Statistics and Computer Applications in Food and Dairy Technology	-	1	1	-	1 0	-	40	50	1
<b>Total</b>				12	6	18	6 0	6 0	2 4 0	24 0	60 0	21
<b>Mandatory Non CGPA Courses</b>												

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<b>Non - CG PA</b>	1 4	UG FD – T306 <b>SECC</b>	Leadership Development	0.5	-	0.5	2 5	-	-	-	25	0.5
	1 5	UG FD – T307 <b>AECC</b>	Environmental Studies – I	0.5	-	0.5	2 5	-	-	-	25	0.5
<b>Total</b>				1	-	1	5 0	-	-	-	50	1
<p style="text-align: center;"><b>Total Credits for Semester III : 22 (T = Theory: 15, P = Practical : 6, Non-CGPA : 1)</b>  <b>CC : Core Course, CCS : Core Course Specialization, DSC : Discipline Specific Course,</b>  <b>DSE : Discipline Specific Elective,</b>  <b>SECC = Skill Enhancement Compulsory Course : 0.5, AECC = Ability Enhancement</b>  <b>Compulsory Course : 0.5,</b>  <b>Total Credits for Semester III CGPA Course = 21 credits</b></p>												

**B.Sc. Food and Dairy Technology Part II, Semester IV**

<b>B.Sc. Food and Dairy Technology Part II, Semester IV (w.e.f. 2023-2024)</b>												
S r · N o	Course Code	Course Title	Teaching Hours/ Week			Marks				Cre dits		
			T	P	To tal	Inter nal		Exter nal			To tal	
						T	P	T	P			
<b>CGPA Theory Courses</b>												
<b>CG PA</b>	1	UG FD – T401 CC	Food Production Technology – I	3	-	3	1 0	-	4 0	-	50	2.5
	2	UG FD – T402 CC	Food Production Technology – II	3	-	3	1 0	-	4 0	-	50	2.5
	3	UG FD – T403 CC	Food Production Technology – III	3	-	3	1 0	-	4 0	-	50	2.5
	4	UG FD – T404 CC	Dairy Production Technology – I	3	-	3	1 0	-	4 0	-	50	2.5
	5	UG FD – T405 CCS	Dairy Production Technology – II	3	-	3	1 0	-	4 0	-	50	2.5
	6	UG FD – T406 DSC	Dairy Production Technology – III	3	-	3	1 0	-	4 0	-	50	2.5
<b>CGPA Practical Courses</b>												
<b>CG PA</b>	7	UG FD – P401 CC	Practicals related to the theory paper - Food Production Technology – I	-	1	1	-	1 0	-	40	50	1
	8	UG FD – P402 CC	Practicals related to the theory paper - Food Production Technology – II	-	1	1	-	1 0	-	40	50	1
	9	UG FD – P403 CC	Practicals related to the theory paper - Food Production Technology – III	-	1	1	-	1 0	-	40	50	1
	10	UG FD – P404 CC	Practicals related to the theory paper - Dairy Production Technology – I	-	1	1	-	1 0	-	40	50	1
	11	UG FD – P405 CCS	Practicals related to the theory paper - Dairy Production Technology – II	-	1	1	-	1 0	-	40	50	1
	12	UG FD – P406 DSC	Practicals related to the theory paper - Dairy Production Technology – III	-	1	1	-	1 0	-	40	50	1
	13	UG FD – P407 PP	Project II	-	1	1	-	1 0	-	10	20	1
	<b>Total</b>				18	7	25	6 0	7 0	2 4 0	25 0	62 0
<b>Mandatory Non CGPA Courses</b>												

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<b>Non - CG PA</b>	1 4	UG FD – T406 <b>SECC</b>	Indian Constitution and Governance	0.5	-	0.5	2 5	-	-	-	25	0.5
	1 5	UG FD – T407 <b>AECC</b>	Environmental Studies – II	0.5	-	0.5	2 5	-	-	-	25	0.5
<b>Total</b>				1	-	1	5 0	-	-	-	50	1
<p style="text-align: center;"><b>Total Credits for Semester IV : 23 (T = Theory: 15, P = Practical : 6, Project : 1, Non- CGPA : 1)</b></p> <p style="text-align: center;"><b>CC : Core Course, CCS : Core Course Specialization, DSC : Discipline Specific Course, DSE : Discipline Specific Elective, , PP : Project</b></p> <p style="text-align: center;"><b>SECC = Skill Enhancement Compulsory Course : 0.5, AECC = Ability Enhancement Compulsory Course : 0.5,</b></p> <p style="text-align: center;"><b>Total Credits for Semester IV CGPA Course = 22 credits</b></p>												

**B.Sc. Food and Dairy Technology Part III, Semester V**

<b>B.Sc. Food and Dairy Technology Part III, Semester V (w.e.f. 2024-2025)</b>												
	S r . N o	Course Code	Course Title	Teaching Hours/ Week			Marks					Cre dits
							Inter nal		Exter nal		To tal	
				T	P	To tal	T	P	T	P		
<b>CGPA Theory Courses</b>												
<b>CG PA</b>	1	UG FD – T501 CCS	Food Production - I	4	-	4	2 0	-	8 0	-	10 0	4
	2	UG FD – T502 DSC	Food Production - II	4	-	4	2 0	-	8 0	-	10 0	4
	3	UG FD – T503 CC	Production in Dairy - I	4	-	4	2 0	-	8 0	-	10 0	4
	4	UG FD – T504 DSE	Production in Dairy - II	4	-	4	2 0	-	8 0	-	10 0	4
	5	UG FD – T505 DSE	Wastewater Technology									
<b>CGPA Practical Courses</b>												
<b>CG PA</b>	6	UG FD – P501 CCS	Practicals related to the theory paper - Food Production - I	-	1	1	-	1 0	-	40	50	1
	7	UG FD – P502 DSC	Practicals related to the theory paper - Food Production - II	-	1	1	-	1 0	-	40	50	1
	8	UG FD – P503 CC	Practicals related to the theory paper - Production in Dairy - I	-	1	1	-	1 0	-	40	50	1
	9	UG FD – P504 DSE	Practicals related to the theory paper - Production in Dairy - II	-	1	1	-	1 0	-	40	50	1
	10	UG FD – P505 DSE	Practicals related to the theory paper - Wastewater Technology									
<b>Total</b>				12	4	16	8 0	4 0	3 2 0	16 0	60 5	20
<b>Mandatory Non CGPA Courses</b>												
<b>Non - CG PA</b>	12	UG FD – T506 SECC	Personal Hygiene and Cleanliness	0.5	-	0.5	2 5	-	-	-	25	0.5
	13	UG FD – T507 AECC	Cyber Security	0.5	-	0.5	2 5	-	-	-	25	0.5
<b>Total</b>				1	-	1	5 0	-	-	-	50	1
<b>Total Credits for Semester V : 21 (T = Theory: 16, P = Practical : 4, Non-CGPA : 1)</b>												



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**CC : Core Course, CCS : Core Course Specialization, DSC : Discipline Specific Course,  
DSE : Discipline Specific Elective,  
SECC = Skill Enhancement Compulsory Course : 0.5, AECC = Ability Enhancement  
Compulsory Course : 0.5,  
Total Credits for Semester V CGPA Course = 20 credits**

**B.Sc. Food and Dairy Technology Part III, Semester VI**

<b>B.Sc. Food and Dairy Technology Part III, Semester VI (w.e.f. 2024-2025)</b>												
S r . N o	Course Code	Course Title	Teaching Hours/ Week			Marks					Cre dits	
			T	P	To tal	Inter nal		Exter nal		To tal		
						T	P	T	P			
<b>CGPA Theory Courses</b>												
<b>CG PA</b>	1	UG FD – T601 CCS	Miscellaneous Food Products	4	-	4	2 0	-	8 0	-	10 0	4
	2	UG FD – T602 DSC	Packaging and Quality Assurance in Food and Dairy Industry	4	-	4	2 0	-	8 0	-	10 0	4
	3	UG FD – T603 CC	Management of Finance in Food and Dairy Industry	4	-	4	2 0	-	8 0	-	10 0	4
	4	UG FD – T604 DSE	Marketing of Food and Dairy Products	4	-	4	2 0	-	8 0	-	10 0	4
	5	UG FD – T605 DSE	Environmental Sciences in Food and Dairy Industry									
<b>CGPA Practical Courses</b>												
<b>CG PA</b>	6	UG FD – P601 CCS	Practicals related to the theory paper - Miscellaneous Food Products	-	1	1	-	1 0	-	40	50	1
	7	UG FD – P602 DSC	Practicals related to the theory paper - Packaging and Quality Assurance in Food and Dairy Industry	-	1	1	-	1 0	-	40	50	1
	8	UG FD – P603 CC	Practicals related to the theory paper - Management of Finance in Food and Dairy Industry	-	1	1	-	1 0	-	40	50	1
	9	UG FD – P604 DSE	Practicals related to the theory paper - Marketing of Food and Dairy Products	-	1	1	-	1 0	-	40	50	1
	10	UG FD – P605 DSE	Practicals related to the theory paper - Environmental Sciences in Food and Dairy Industry									
	11	UG FD – P606 PP	Project III	-	1	1	-	1 5	-	10	25	0.5
<b>Total</b>				12	5	17	8 0	5 5	3 2 0	17 0	62 5	21
<b>Mandatory Non CGPA Courses</b>												
<b>Non - CG</b>	1 2	UG FD – T606 SECC	Human Rights and Human Values	0.5	-	0.5	2 5	-	-	-	25	0.5

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<b>PA</b>	1 3	UG FD – T607 AECC	Food and Dairy Data Care Management	0.5	-	0.5	2 5	-	-	-	25	0.5
<b>Total</b>				1	-	1	5 0	-	-	-	50	1
<p><b>Total Credits for Semester VI : 22 (T = Theory: 16, P = Practical : 4, Project : 1, Non-CGPA : 1)</b>  <b>CC : Core Course, CCS : Core Course Specialization, DSC : Discipline Specific Course, DSE : Discipline Specific Elective, PP : Project</b>  <b>SECC = Skill Enhancement Compulsory Course : 0.5, AECC = Ability Enhancement Compulsory Course : 0.5,</b>  <b>Total Credits for Semester VI CGPA Course = 21 credits</b></p>												

**UG HM - T101: Fundamentals of Microbial and Biological World**

**3-Credits-60-hours**

<b>Unit I</b>	<b>20 Hrs</b>	<p><b>History – Three centuries of Microbiology</b></p> <p><b>A. Development of Microbiology as a discipline:-</b> Discovery of microscope and microorganisms (Antony Van Leeuwenhoek and Robert Hooke), abiogenesis versus biogenesis (Aristotle’s notion about spontaneous generation, Francesco Redi’s experiment, Louis Pasteur and Tyndall’s experiments)</p> <p><b>B. Golden era of Microbiology –</b> Contributions of Louis Pasteur (Fermentation, Rabies vaccine, pasteurization and cholera vaccine – Foul cholera experiment), Robert Koch (Koch’s postulates, germ theory of diseases, Tuberculosis and Cholera – isolation and staining techniques of causative agent, pure culture techniques), Ferdinand Cohn (Endospore Discovery), discovery of viruses (TMV- Ivanowsky and bacteriophages- deHerrale), Rivar’s postulates, Contributions of Joseph Lister (Antiseptic Surgery), Paul Ehrlich (chemotherapy), Elie Metchnikoff (Phagocytosis), Edward Jenner (Vaccination), Alexander Flemming (Penicillin) and Selman Waksman (Streptomycin) in the establishment of fields of medical microbiology and immunology. Contributions of Martinus W. Beijerinck (Enrichment culture technique, Rhizobium), Sergei. N. Winogradsky (Nitrogen Fixation, azatobacter and Chemolithotrophy) in the development of fields of soil microbiology.</p> <p><b>C. Modern era of Microbiology</b> Prokaryotic and Eukaryotic Classification – Three domain and five domain systems, Carl Woese classification based on 16S rRNA gene sequencing. Significance and applications of human microbiome, nanoFood and Dairy, space microbiology, geomicrobiology and r-DNA technology Nobel Laureates in Life Sciences of 21<sup>st</sup> Century</p>
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<b>Unit II</b>	<b>15 Hrs</b>	<p><b>Types of Microorganisms and their differentiating features</b></p> <p><b>A) Cellular forms</b> – Prokaryotic and eukaryotic          Bacteria (Eubacteria, archaebacteria, Rickettsia, Mycoplasma and Actinomycetes)          Protozoa , Fungi, Algae</p> <p><b>B) Acellular Forms</b> – Viruses, Viroids, Prions, Virusoid</p>
<b>Unit III</b>	<b>25 Hrs</b>	<p><b>Beneficial and harmful effects of microorganisms in various fields of Microbiology, Food and Dairy and Environmental Sciences:</b></p> <p>a) Medical Microbiology (Enlist diseases caused by various microorganisms, vaccines and antibiotics)</p> <p>b) Immunology (Normal Flora, Immune Sera, Three lines of defenses)</p> <p>c) Food and Dairy Microbiology (Food spoilage, food borne diseases, prebiotics, probiotics and fermented foods)</p> <p>d) Industrial microbiology (Microorganisms producing antibiotics, enzymes, growth factors, solvents and SCP; contaminants in industry– bacteria, fungi and phages)</p> <p>e) Agricultural Microbiology (Enlist plant diseases, biofertilizers, plant growth promoters and biocontrol agents)</p> <p>f) Space microbiology (Space microbes as a tool to study origin of life on the earth)</p> <p>g) Geomicrobiology (Metal leaching from ores)</p> <p>h) NanoFood and Dairy (Production of nanoparticles using microorganisms)</p>

**UG HM - P101: Practical course based on theory paper**

**Fundamentals of Microbial and Biological World**

**30 hrs.**

1	Introduction, operation, precautions and use of common laboratory instruments used in life sciences [Incubator, Hot air oven, Autoclave, Colorimeter, Centrifuge, Laminar air flow, pH meter, Digital balance, Microscopes, Anaerobic jar, Colony counter, Seitz Filter, Distillation Unit, Membrane Filter]	06 Hrs.
2	Learning basic techniques in life science laboratory [Washing, plugging and wrapping of glassware, biological waste disposal, aseptic transfer techniques – broth, plate, slant and butt transfers]	04 Hrs.
3	Observation of motility in bacteria by hanging drop/ swarming growth method	02 Hrs.
4	Checking efficiency of chemical disinfectants - Phenol coefficient by Rideal- Walker method	8 Hrs.
5	Special staining techniques- Cell wall (Chance's method), flagella (Bailey's method/Leifson's method), acid fast staining (permanent slide)	10 Hrs

<b>Unit I</b>	<b>6 Hrs</b>	<b>Measurements</b> <ul style="list-style-type: none"> <li>Physical quantities, fundamental and derived units, system of units, order of magnitude</li> <li>Length: radius of proton to astronomical distances</li> <li>Mass: atomic mass unit to mass of earth</li> <li>Time: fast elementary particle to age of earth</li> <li>Amount of substance, luminous intensity, interconversions of units</li> </ul>
<b>Unit II</b>	<b>7 Hrs</b>	<b>Introduction to biophysics</b> <ul style="list-style-type: none"> <li>Scope and definition of biophysics, biophysics at macroscopic, microscopic and molecular level.</li> <li>Biophysical properties: Surface tension, adsorption, diffusion, osmosis, dialysis, wetting and colloids</li> </ul>
<b>Unit III</b>	<b>10 Hrs</b>	<b>Fluid Mechanics: (5)</b> <ul style="list-style-type: none"> <li>Fluids: definition, pressure, density, variation of pressure with depth in a fluid at rest,</li> <li>Measurement of pressure- Various units of pressure and their interconversion, streamline and turbulent flow</li> <li>Equation of Continuity, Poiseuille's equation, Reynold's number, flow of liquids through capillaries, viscosity, Newton's law of viscosity, coefficient of viscosity, Ostwald's viscometer, Relevance to life Science, Bernoulli's theorem and its applications, methods of measurement of viscosity</li> </ul>
<b>Unit IV</b>	<b>8 Hrs</b>	<b>Surface Tension &amp; Surface Energy</b> <ul style="list-style-type: none"> <li>Cohesive and adhesive forces, Capillary action, angle of contact, wettability, measurement of surface tension by capillary rise, Jaeger's and Quincke's method, factor affecting surface tension, applications, relevance to life sciences</li> </ul>
<b>Unit V</b>	<b>12 Hrs</b>	<b>Waves &amp; Oscillations</b> <ul style="list-style-type: none"> <li>Difference between waves and oscillations, Types of waves (Transverse &amp; Longitudinal), Reflection of waves, Principle of superposition of waves, standing &amp; travelling waves, Sound waves as pressure waves, Audible ultrasonic &amp; infrasonic waves, characteristics of sound waves, vibration systems and source of sound, beats, Doppler's effect, Applications in life sciences, measurement of sound, decibel scale (dB).</li> </ul>
<b>Unit VI</b>	<b>7 Hrs</b>	<b>Geometrical Optics</b> <ul style="list-style-type: none"> <li>Reflection, Refraction, Snell's Law, types of lenses, combinational lenses, radius of curvature, focal length, lens maker equation.</li> </ul>
<b>Unit VII</b>	<b>10 Hrs</b>	<b>Radioactivity:</b> <ul style="list-style-type: none"> <li>Nucleus: Properties – size, shape, charge distribution, spin and purity binding and empirical mass formula, nuclear stability and radioactive decay, nuclear forces, nuclear models (Liquid drop &amp; Shell model), radioactive nucleus Nuclear Radiations &amp; their properties, Alpha, Beta &amp; Gamma, half life, Physical &amp; biological handling of alpha &amp; beta emitting isotopes, UV and X-rays – properties, X-ray spectrum, Braig's law and</li> </ul>

		applications GM Counter – Principle, construction & working
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**UG HM - P102: Practicals related to theory paper Fundamentals of Physics  
and Biophysics for Biologists**

**30 hrs.**

1	Study of Vernier callipers & micrometer screw gauge	03 hrs.
2	To Study the components & working of travelling microscope	04 hrs.
3	Surface tension measurement using Jaeger's method/ Soap bubble method	03 hrs.
4	To Study plane diffraction grating	04 hrs.
5	Special staining techniques- Cell wall (Chance's method), flagella (Bailey's method/Leifson's method), acid fast staining (permanent slide)	04 hrs
6	Study the process of osmosis	02 hrs
7	Determination of diffusion pressure deficit using potato tuber	02 hrs
8	Precipitation & Dialysis	02 hrs
9	Working of GM counter	02 hrs
10	Sonometer	02 hrs
11	Determine surface tension of liquids	02 hrs



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**B. Sc. Part I Semester – I**

**UG HM - T103 Fundamentals of Chemistry for Biologists**

**3-Credits-60-h**

<b>Unit I</b>	<b>10 Hrs</b>	<p><b>Atomic Structure</b>  Historical background, electronic structure of atom, JJ Thomson and Rutherford model, Bohr's Model and its postulates, atomic and molecular orbitals, four quantum numbers, shapes of atomic orbitals, selection rules to find out electronic configuration of elements, Plank's quantum theory, Wave particle duality, Uncertainty principle, Pauly's exclusion principle, Ionisation Potential, electronegativity, electron affinity</p>
<b>Unit II</b>	<b>6 Hrs</b>	<p><b>Molecules</b>  Diatomic molecules, valance bond theory, VSEPR theory, hybridization involving s, p, d orbitals (<math>sp</math>, <math>sp^2</math>, <math>sp^3</math>, <math>dsp^2</math>, <math>sp^2d</math>, <math>sp^3d^2</math>), homo and heteronuclear diatomic molecules, bond order, magnetic properties</p>
<b>Unit III</b>	<b>6 Hrs</b>	<p><b>. Chemical Bonding</b>  Types of bonds: covalent, coordinate, metallic, ionic, hydrogen bonding, inter and intramolecular hydrogen bonding, dipole-dipole, dipole induced dipole interaction, structure of water molecule, oxidation state, hydrophobic and hydrophilic interactions</p>
<b>Unit IV</b>	<b>10 Hrs</b>	<p><b>Basics of Organic and Stereochemistry and mechanisms</b></p> <ul style="list-style-type: none"> <li>• IUPAC nomenclature,</li> <li>• reactions of functional groups : alkane, alkene, alkyne, alcohol, amine, alkyl halide, ether,</li> <li>• organic reactions : oxidation, reduction, elimination, addition, substitution (electrophilic/ nucleophilic), inductive, mesomeric and electrometric effects, reactive intermediates – carbonations, carbon ion, free radicals, carbines, Arynes and Nytrins</li> <li>• Conformations, configurations, isomerism (structural and stereo isomers), enantiomers, diastereoisomers, chiral centers, geometric isomers, optical isomerism</li> <li>• Newman's and Fisher projection formulae, epimers, anomers, furanose, and pyranose forms, free radical reactions</li> </ul>
<b>Unit V</b>	<b>6 Hrs</b>	<p><b>. Ionic Equilibrium</b></p> <ul style="list-style-type: none"> <li>• pH, buffer, equilibrium constant, common ion effect, Le Chatelier's principle, acids and bases, strength of acids and bases, dissociation constant, pH, pK values, solubility product, acid-base titrations, indicators used in titration, titration curves, Bronstied-Lowery theory, Levis theory, Acid-base concept in non gaseous solvents, Soft hard acid bases (SHAB) concept</li> <li>• Ionic product, condition for precipitation, colligative properties of solutions</li> <li>• Handerson – Hasselbalch equation and related problems, osmosis, law of osmotic pressure and its measurement, determination of molecular</li> </ul>

		<p>weight from osmotic pressure</p> <ul style="list-style-type: none"> <li>• Properties of water, water as reactant, interactions of biomolecules with water</li> </ul>
<b>Unit VI</b>	<b>7 Hrs</b>	<p><b>Chemical Kinetics</b></p> <ul style="list-style-type: none"> <li>• Rates of reactions, order - zero, first and second order reactions and molecularity</li> <li>• Differential and integrated rate equation, methods of determining order of reactions, catalysis and elementary enzyme reactions</li> <li>• Half- life periods, Arrhenius equation, collision theory of reaction rate, temperature dependent reaction rates</li> </ul>
<b>Unit VII</b>	<b>10 Hrs</b>	<p><b>Thermodynamics</b></p> <p>Introduction, types of system, intensive and extensive properties, equilibrium and non-equilibrium states, reversible and irreversible processes, laws of thermodynamics, internal energy, enthalpy, entropy – basic concept, physical significance, principle of increase in natural processes, endothermic and exothermic reactions, free energy and work, Gibb's Helmholtz equations, Isothermal and adiabatic relation, work done during isothermal and adiabatic changes, Carnot's engine and Carnot's cycle and its efficiency, Practical cycle used in internal combustion in engine (diesel engine)</p>
<b>Unit VIII</b>	<b>5 Hrs</b>	<p><b>Basics of Mole Concept</b></p> <ul style="list-style-type: none"> <li>• Mole concept, determination of molecular weight by gram molecular volume relationship, problems based on mole concept, solutions, colligative properties</li> <li>• Methods of expressing concentrations, strength, normality, molarity and molality, ppm</li> <li>• Volumetric experiments – acidometry, alkalometry, permanganometry, dichrometry, iodometry</li> </ul>

1	<b>Titration</b> a. To study acid – base titration by indicator and conductivity meter b. To determine alkali content on antacid tablet using HCl	04 hrs.
2	<b>Chemical kinetics</b> To study kinetics of ester's hydrolysis	03 hrs.
3	<b>Thermochemistry</b> To determine enthalpy and entropy change of a reaction e.g.(1) $2\text{FeCl}_3 + 3\text{Mg} = 2\text{Fe} + 3\text{MgCl}_2$ Activation energy for an acid catalyzed hydrolysis of methyl acetate	03 hrs.
4	<b>Hardness of water</b> To estimate hardness of water by using EDTA	02 hrs.
5	<b>Qualitative analysis</b> To perform qualitative test for hydrocarbons, alcohols, aldehydes, ketones, aniline and amide	06 hrs
6	<b>pH meter</b> To determine pK value of given weak acid by pH meter titration with strong base	02 hrs
7	<b>Biochemical calculation</b> Preparation of solutions and buffers (Normality, Molarity, molality, parts per million - ppm, weight by volume - w/v, volume by volume - v/v, percent - %, atomic weight, molecular weight, equivalent weight) Preparation of dilute solution from given stock solution (concentrated saline citrate, dilute saline citrate, normal/standard saline citrate)	04 hrs
8	To study different conformation of biomolecules using models	02 hrs
9	Organic preparations – Pthalimide, Methyl Salicylate	02 hrs
10	Sonometer	02 hrs
11	Inorganic preparations – Hexamine Nickel (II) chloride	02 hrs

<b>Unit I</b>	<b>12 Hrs</b>	<b>Introduction to plant world and classification (Plant Diversity)</b> <ul style="list-style-type: none"> <li>➤ General and unique features of plants</li> <li>➤ Principles, aims, objectives and outline of plant classification with examples</li> <li>➤ A general account of different groups and their characters with one example each of <ul style="list-style-type: none"> <li>○ Thallophytes (Algae, Fungi and Lichens)</li> <li>○ Bryophytes</li> <li>○ Pteridophytes</li> <li>○ Gymnosperms</li> <li>○ Angiosperms (Dicot and Monocot)</li> </ul> </li> </ul>
<b>Unit II</b>	<b>18 Hrs</b>	<b>Structure and organization of plant body</b> <ul style="list-style-type: none"> <li>➤ Structure of plant cell, characteristic feature and cell wall</li> <li>➤ Morphology &amp; modifications of plant organs <ul style="list-style-type: none"> <li>○ Vegetative plant organs – Stem, Leaf and Root</li> <li>○ Reproductive plant organs – Flower and Types of Inflorescence</li> </ul> </li> <li>➤ Plant tissues and tissue systems <ul style="list-style-type: none"> <li>○ Meristematic tissue and its type</li> <li>○ Permanent tissue - Simple and Complex</li> </ul> </li> <li>➤ Primary structure of shoot, root &amp; leaf</li> <li>➤ Secondary growth, growth rings formation: cambium and its activities, periderm- cork cambium, secondary cortex and cork</li> </ul>
<b>Unit III</b>	<b>20 Hrs</b>	<b>Introduction to Kingdom Animalia</b> <ul style="list-style-type: none"> <li>➤ Outline classification of non-chordates with examples <ul style="list-style-type: none"> <li>○ General characters and classification up to classes of phylum Porifera, Cnidaria, Platyhelminthes, Nematelminthes, Annelida, Arthropoda, Mollusca, Echinodermata and Hemicordata</li> </ul> </li> <li>➤ Outline classification of chordates with examples <ul style="list-style-type: none"> <li>○ General characters and classification up to classes of phylum Protochordates, Agnatha, Pisces, Amphibia, Reptiles, Aves and Mammals</li> </ul> </li> </ul>
<b>Unit IV</b>	<b>10 Hrs</b>	<b>Animal Tissues (Histology)</b> <ul style="list-style-type: none"> <li>➤ Structure, location, classification and functions of animal tissues <ul style="list-style-type: none"> <li>○ epithelial tissue</li> <li>○ connective tissue</li> <li>○ muscular tissue</li> <li>○ nervous tissue</li> </ul> </li> <li>➤ Bone and Cartilage - structure and types</li> </ul>

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**UG HM - P104: Practical in Biosciences – Botany and Zoology**

**30 hrs.**

1	Study of - Thallophytes (Algae, Fungi and Lichens), Bryophytes, Pteridophytes, Gymnosperms with one example each	04 hrs.
2	Study of morphological parameters of Angiosperms (Dicot and Monocot)	03 hrs.
3	Study on anatomy of root, stem, leaf of monocot and dicot plants	04 hrs.
4	Study of Paramecium – morphology, reproduction, binary fission, conjugation	04 hrs.
5	Study of phylum – Porifera, Cnidaria, Platyhelminthes, Nematelminthes, Annelida, Arthropoda, Mollusca, Echinodermata, Protochordates, Agnatha, Pisces, Amphibia, Reptiles, Aves and Mammals with one example each (specimen)	10 hrs
6	Study of Drosophila: characters, sexual dimorphism – eye & wing mutations, life cycle, culturing of Drosophila	03 hrs
7	Staining of Animal and Plant Cells	02 hrs

**B. Sc. Part I Semester – I**

**UG HM - T105: Basics of bacteriology, Virology & Rickettsiology**

**3-Credits-60-h**

<b>Unit I</b>	<b>20 Hrs</b>	<b>Bacteriology</b> Types of bacteria as per their carbon and energy requirements (nutritional classification), advanced classification of bacteria with example using G + C content, DNA –RNA hybridisation, 16 S rRNA gene sequencing & fatty acid lipid profile
<b>Unit II</b>	<b>20 Hrs</b>	<b>Virology</b> Discovery, nature of viruses, types of viruses, outline classification with example, structure of viruses <ul style="list-style-type: none"><li>• <b>Bacteriophages</b> -T4 cycle &amp; cultivation (Coliphages)</li><li>• <b>Animal Viruses</b> – Types, cultivation, AIDS, Swine Flu, Dengue, Corona viruses – Life cycle &amp; control</li><li>• <b>Plant viruses</b> – Outline classification with examples, life cycle, and control mechanisms.</li><li>• Applications of viral genomes in Food and Dairy, microbiology &amp; Environmental sciences</li><li>• Viroids, prion and virusoides</li></ul>
<b>Unit III</b>	<b>20 Hrs</b>	<b>Rickettsiology</b> Unique features of Rickettsia, Outline Classification, cultivation, significance, control measures Vaccines in Rickettsial infections <i>Coxiella burnetii</i> , <i>Chlamydia</i> & <i>Mycoplasmas</i> – General characteristics & significance

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**UG HM - P105: Practical in Basics of bacteriology, Virology & Rickettsiology**

**30 hrs.**

1	Isolation of pigment producing yeast / bacteria from nature	06 hrs.
2	Isolation & cultivation of autotrophs and heterotrophs	06 hrs.
3	Isolation & titration of bacteriophages (Coliphages) from sewage	06 hrs.
4	Inoculation of Viruses - Egg inoculation technique & cultivation of viruses	06 hrs.
5	Animal viruses - AIDS, Swine Flu, Dengue, Corona, Chikungunia (chart/ animation)	02 hrs
6	Plant Viruses - TMV / Leaf curl virus (chart/ animation)	02 hrs
7	Rickettsia- life cycle study (Photos / Demonstration/ Charts/ Digital/ Animation)	02 hrs

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**B. Sc. Part I Semester – I**

**UG HM - T106: Fundamentals of Mycology, Phycology & Protozoology**

**3-Credits-60-h**

<b>Unit I</b>	<b>20 Hrs</b>	<b>Mycology – Yeasts and molds</b> <ul style="list-style-type: none"><li>- Outline classification, characteristics, structure and reproduction</li><li>- Cultivation of yeasts and molds</li><li>- Life cycle of yeasts and molds</li><li>- Biological and economic importance</li><li>- Important features and significance of slime molds, myxomycetes, mycorrhiza and mushrooms</li></ul>
<b>Unit II</b>	<b>20 Hrs</b>	<b>Phycology – Algae</b> <ul style="list-style-type: none"><li>- Outline classification, morphological characteristics, cultivation, reproduction and significance</li><li>- Characteristics of algae, pigments, major groups – an overview</li><li>- Biological, medical and economic importance of algae</li><li>- Differences between algae and cyanobacteria</li><li>- Examples of toxic algal forms in drinking water</li></ul>
<b>Unit III</b>	<b>20 Hrs</b>	<b>Protozoology – Protozoa</b> <ul style="list-style-type: none"><li>- Outline classification, morphological characteristics, cultivation, reproduction and significance</li><li>- Major categories of protozoa based on motility and reproduction</li><li>- Medically important protozoa</li><li>- Life cycle of <i>Entamoeba histolytica</i></li></ul>



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**UG HM - P106: Practical Fundamentals of Mycology, Phycology & Protozoology**

**30 hrs.**

1	Isolation and cultivation of algae/ cyanobacteria [Spirulina/Chlorella/Scytonemia]	06 hrs.
2	SCP – Extraction from Spirulina/ Study of mushroom/ Study of lichens	06 hrs.
3	Isolation & titration of bacteriophages (Coliphages) from sewage	06 hrs.
4	Isolation of wine yeasts from spoiled pomegranate and preparation of wine	06 hrs.
5	Isolation and cultivation of <i>Aspergillus niger</i> [from onion]/ <i>Penicillium/ Mucor/ Rhizopus/ Fusarium spp.</i> from soil	02 hrs
6	Plant Viruses - TMV / Leaf curl virus (chart/ animation)	02 hrs
7	Detection, isolation [single cell isolation technique] and cultivation of protozoa from water bodies, [Zooplanktons/ Paramecium/Amoeba/Euglena/ Vorticella studies from water]	02 hrs

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**B. Sc. Part I Semester – I**

**UG HM - T107: Introduction to the world of amazing microorganism**

**3-Credits-60-h**

<b>Unit I</b>	<b>3 Hrs</b>	Autotrophic microorganisms- occurrence, characteristics, mechanism, energetics, significance & examples; Biocorrosion and Bioleaching ( <i>Thiobacillus</i> )
<b>Unit II</b>	<b>6 Hrs</b>	Bioluminescent forms- Luminescence in nature, bioluminescence, bioluminescent bacteria & fungi- characteristics, occurrence, mechanism, energetics & significance in nature
<b>Unit III</b>	<b>6 Hrs</b>	Magnetostatic forms- Magnetotactic bacteria occurrence, mechanism, mechanism of magnet axis, their role in detection of exotic (in space) life, significance in nature; Astrobiology (introduction to space environment and space microbiology)
<b>Unit IV</b>	<b>6 Hrs</b>	Extremophiles- Psychrophiles, acidophiles, xerophiles, barophiles, halophiles, radiophiles, thermophiles, basophiles, piezophiles, osmophiles - occurrence, characteristics, mechanism of survival, energetics, significance & examples
<b>Unit V</b>	<b>6 Hrs</b>	Bdellovibrio forms- examples, occurrence, characteristics, nature of parasitism, hmechanism & significance
<b>Unit VI</b>	<b>6 Hrs</b>	Bacteria visible by naked eye (largest bacteria) - examples, occurrence & significance
<b>Unit VII</b>	<b>6 Hrs</b>	Obligate intracellular parasitic microorganisms - examples - <i>Rickettsia</i> , viruses- (animal viruses, plant viruses, bacterial viruses)
<b>Unit-VIII</b>	<b>6 Hrs</b>	Actinomycetes & Myxobacteria
<b>Unit-IX</b>	<b>6 Hrs</b>	Unculturable Microorganisms (metagenomic study) - <i>Mycobacterium leprae</i> - The organism not following Koch's postulates, their significance in nature
<b>Unit-X</b>	<b>3 Hrs</b>	Nitrogen fixing bacteria in nature, examples, mechanism and significance
<b>Unit XI</b>	<b>6 Hrs</b>	Aromatic Compounds, plastic, Cyanide degrading microorganism – <i>Pseudomonas putida</i> (Anand Chakravorty)

**UG HM - P107: Practical Introduction to the world of amazing microorganism**

**30 hrs.**

1	Isolation, cultivation & characterization of bioluminescent bacteria	06 hrs.
2	Isolation, cultivation & characterization of Magnetotactic bacteria (Optional)	06 hrs.
3	Isolation & cultivation of Actinomycetes/Myxobacteria	04 hrs.
4	Isolation, Cultivation and Characterization of Bdellovibrio forms	06 hrs.
5	Isolation of bacteria degrading microplastic/ aromatic compounds/ cyanide	02 hrs
6	Isolation of <i>Azotobacter/Rhizobium</i> (Optional)	02 hrs
7	Isolation cultivation & characterization of Extremophiles – Psychrophiles/ Thermophiles/ Barophiles/ radiophiles/ basophiles/ acidophiles/ xerophiles/ piezophiles/ halophiles/ osmophiles	02 hrs
8	Slide of <i>Mycobacterium leprae</i> - acid fast stains, demonstration (Optional)	02 hrs

**B. Sc. Part I Semester – I**

**UG HM - T108: Basics Tools and Techniques in Microbiology, Food and Dairy and  
Environmental Sciences**

**3-Credits-60-h**

<b>Unit I</b>	12 Hrs	<p>Safety in Life Sciences laboratory</p> <ul style="list-style-type: none"> <li>• Means of laboratory infections</li> <li>• Potentially hazardous procedures</li> <li>• Responsibility</li> <li>• Risk assessment</li> <li>• Restricted access</li> <li>• Safety equipments and measures</li> <li>• Immunization and medical records</li> <li>• Training of personnel</li> <li>• Laboratory procedures (SOPs)</li> <li>• Levels of containments</li> </ul>
<b>Unit II</b>	12 Hrs	<p>Microscopy</p> <p>A. Bright field microscopy:</p> <ol style="list-style-type: none"> <li>a. Electromagnetic spectrum of light</li> <li>b. Simple and compound microscope - working of and ray diagram; concepts of magnification, numerical aperture and resolving power. Types functions of - eyepieces and objectives; aberrations in lenses - spherical, chromatic, comma and astigmatism</li> <li>c. Phase contrast microscopy – mechanism and applications</li> <li>d. Fluorescence Microscopy – mechanism and applications</li> <li>e. Electron Microscopy – Basic principle, mechanism, TEM, SEM, STM and their applications</li> </ol> <p>- B. Dark field microscopy: Mechanism and applications</p>
<b>Unit III</b>	4 Hrs	Chromatography – Paper and TLC, theory, instrument and applications
<b>Unit IV</b>	12 Hrs	<p>Observation of cells:</p> <p>A. Stains and staining techniques</p> <ol style="list-style-type: none"> <li>a. Definition of Stain; Types of stains (Basic, Acidic and Neutral), Properties and role of Fixatives, Mordants, Decolourisers and Accentuators</li> <li>b. Staining procedures for bacteria – Monochrome (Simple) staining and Negative (Relief) staining</li> <li>c. Differential staining - Gram staining and Acid-fast staining – mechanism and procedure</li> <li>d. Special staining- mechanism and procedure - Capsule, Cell wall, Endospore, Flagella, Nuclear material, Lipid granules,</li> </ol>

		<p style="text-align: center;">metachromatic granules</p> <p style="text-align: center;">e. staining of animal and plant cells</p> <p style="text-align: center;">f. staining of algae, protozoa and fungi</p> <p>B. Unstained preparations – wet mount and hanging drop techniques of bacteria, yeasts, molds, algae and protozoa</p>
<b>Unit V</b>	<b>20 Hrs</b>	<p><b>Control of Microorganisms</b></p> <p>a. Definitions of frequently used terms – sterilization, disinfection, antiseptic, antiseptis, germicide, microbiostasis, sanitization, bactericide, Fungicide, viruside, sporicide, fundamentals of control, conditions influencing effectivity of antimicrobial agent, factors affecting death rate</p> <p>b. Physical agents used to control microorganisms –</p> <ul style="list-style-type: none"> <li>• Heat - Dry and Moist; Radiations-Ionizing (X-ray, gamma and cathode) and Non-ionizing (UV rays); filtration- depth filters and membrane filters (cellulose acetate and polycarbonate filters, plastic – Teflon and Nylon), low and high temperature, osmotic pressure, desiccation, Sound waves – Ultrasonication</li> <li>• Checking the efficacy of sterilization – biological and chemical indicators</li> </ul> <p>c. Chemical agents used to control microorganisms and their mode of action and applications–</p> <ul style="list-style-type: none"> <li>• Characteristics of an ideal disinfectant</li> <li>• Aldehydes, Halogens, Quaternary ammonium compounds, Phenol and Phenolics, peroxigens</li> <li>• Heavy metals (Cu, Hg, Ag), alcohols, dyes, surface active agents, detergents, gaseous agents – ethylene oxide, beta propiolactone, formaldehyde, glutardaldehyde, clorhexidine and benzolkonium chloride</li> <li>• Checking efficiency of disinfectant – phenol coefficient (Rideal-Walker method)</li> <li>• Chemotherapeutic agents (enlist) and their site of action</li> </ul>

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**UG HM - P108: Practical Basics Tools and Techniques in Microbiology, Food and  
Dairy and Environmental Sciences**

**30 hrs.**

1	a) Safety measures and good laboratory practices in the laboratory b) Preparation of SOPs for the instruments c) Introduction and use of common laboratory glass wares	05 hrs.
2	Construction, working and care of compound microscope	04 hrs.
3	Basic staining techniques- Monochrome, Negative and Gram's staining, Acid-fast staining (demo slide)	10 hrs.
4	Special staining techniques- Endospore, Capsule, Lipid granules, Nuclear material, Metachromatic granules, Flagella	11 hrs.

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**B. Sc. Part I Semester – I**

**UG HM - T109SECC: Yoga and Meditation**

**3-Credits-15-h**

<b>Unit I</b>	3 Hrs	Introduction, Meaning, definition, Objectives; Introduction to Ashtangyoga; Performing Yogabhyasa
<b>Unit II</b>	2 Hrs	<b>Suryanamaskar:</b> Introduction, Postures, Benefits and practice
<b>Unit III</b>	7 Hrs	<b>Asanas</b> Vajrasan, Padmasan, Vakrasan, UttanPadmasan, Pawanmuktasan, Shavasana, Bhujangasan, Shalabhasan, Makrasan, Tadasan, Verasan, ArdhaChakrasan- Introduction, Postures, Benefits and practice.
<b>Unit IV</b>	3 Hrs	<b>Pranayamas</b> AnulomVilom, Bhramari, Kapalbhathi and Bhasrika; Omkar Sadhana, Prayer and Guruvandana
<b>Unit V</b>	2 Hrs	<b>Using a Dictionary:</b> Definition of the dictionary, types of dictionaries, information in the dictionary, use of a dictionary
<b>Unit VI</b>	2 Hrs	<b>Use of good English:</b> Noun, pronoun, adjective, verb, adverb, conjunction, preposition, interjection, the article, tenses, spelling, use, and misuse of words, abbreviations, active and passive voice, punctuation, remove 'too'.
<b>Unit VII</b>	2 Hrs	<b>Phonology:</b> Pronunciation of vowels and consonants in English
<b>Unit VIII</b>	1 Hrs	Public speaking in English and oral presentation in English.

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**B. Sc. Part I Semester – I**

**UG HM - T110SECC: Spoken English**

**3-Credits-15-h**

<b>Unit I</b>	<b>1 Hr</b>	<b>Language:</b> English as a foreign language
<b>Unit II</b>	<b>3 Hrs</b>	<b>Writing English:</b> Sentence structure, Essay composition, Summary writing, precise writing and comprehension
<b>Unit III</b>	<b>2 Hrs</b>	<b>Reading English:</b> Importance of reading, the process, and mechanics of reading, Intensive and extensive reading: Rapid reading, making notes as you read, writing book review.
<b>Unit IV</b>	<b>2 Hrs</b>	<b>Use of Vocabulary:</b> Meaning of words, precise usages, synonyms and antonyms, technical terms, context, superfluous words
<b>Unit V</b>	<b>2 Hrs</b>	<b>Using a Dictionary:</b> Definition of the dictionary, types of dictionaries, information in the dictionary, use of a dictionary
<b>Unit VI</b>	<b>2 Hrs</b>	<b>Use of good English:</b> Noun, pronoun, adjective, verb, adverb, conjunction, preposition, interjection, the article, tenses, spelling, use, and misuse of words, abbreviations, active and passive voice, punctuation, remove 'too'.
<b>Unit VII</b>	<b>2 Hrs</b>	<b>Phonology:</b> Pronunciation of vowels and consonants in English
<b>Unit VIII</b>	<b>1 Hrs</b>	Public speaking in English and oral presentation in English.



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**F.Y. B.Sc. Semester II**

**UG HM – T201: Fundamentals of Cell Biology & Physiology**

<b>Unit I</b>	<b>3 Hrs</b>	<b>Introduction to cell:</b> Discovery of cell, cell theory – Definition, three assumptions of cell theory, exceptions, organismal theory, protoplasm theory.
<b>Unit II</b>	<b>4 Hrs</b>	<b>Organization of Prokaryotic cells :</b> size (Micrometry), shape & arrangement of bacterial cells, Structure of typical bacterial cells, Structure & functions of cell wall & cell membrane (Fluid Mosaic Model), composition & functions of capsule, slime layer, flagella, Pili, fimbriae, Cytoplasmic matrices – inclusion bodies, magnetosomes, ribosomes, gas vacuoles, metachromatic granules, Carboxysomes, PHB granules, endospores, Nucleoid & plasmids
<b>Unit III</b>	<b>12 Hrs</b>	<b>Eukaryotic cell structure</b> – Micrometry (Plant & animal cell), Overview of <ul style="list-style-type: none"> <li>▪ eukaryotic cell structure, plasma membrane &amp; membrane structure. Cytoplasmic matrix, microfilaments, intermediate filaments &amp; microtubules</li> <li>▪ <b>Organelles of biosynthesis</b> – Secretary &amp; endocytic pathways – Endoplasmic Reticulum &amp; Golgi apparatus, Definition of Lysosome, Endocytosis, phagocytosis, autophagy &amp; proteosome</li> <li>▪ Eukaryotic Ribosomes, Peroxisomes, Mitochondria, Chloroplast (plastids), Nucleus (Introduction, morphology, occurrence, shape, size, number, position, ultra structure of nucleus, nuclear membrane, nucleoplasma, nucleopore complex, nucleolus, chromosomes – euchromatin &amp; hetero chromatin chromosome number, size, general structure &amp; nomenclature, organization of nucleus, specialized chromosomes - polytene &amp; lampbrush)</li> <li>▪ External cell covering – Cilia &amp; flagella</li> <li>▪ Comparison of prokaryotic &amp; eukaryotic cells</li> </ul>
<b>Unit IV</b>	<b>10 Hrs</b>	<b>Cell membrane &amp; membrane transport :</b> Types of membrane transport – Passive transports – simple diffusion, facilitated diffusion, osmosis, Active transport – Primary & secondary transport, Na –pump, Na <sup>+</sup> - K <sup>+</sup> ATPase pump, bulk transport, endocytosis & exocytosis.
<b>Unit V</b>	<b>5 Hrs</b>	<b>Cell cycle</b> Introduction, phases & check prints – cell division in microorganism & plant, animals (Mitosis & Meiosis) – G <sub>0</sub> , G <sub>1</sub> , G <sub>2</sub> & M phases & significance
<b>Unit VI</b>	<b>10 Hrs</b>	<b>Cell Signalling</b> Signalling molecules, Signalling receptors (cell surface receptors), =autocrine, syncrine & paracrine signalling G-protein signalling & calcium signalling, membrane junctions
<b>Unit VII</b>	<b>6 Hrs</b>	<b>Cell death</b> Aging, Theories of aging, apoptosis & necrosis, neoplasia, autophagy, ferroptosis & pyroptosis
<b>Unit VIII</b>	<b>10 Hrs</b>	Diseases associated with lysosomes (Tay Sachs disease), Peroxysomes (Zell Wager syndrome), Mitochondria (Leber Hereditary Optic Neuropathy -LHON & Mitochondrial encephalomyopathy, lactic acidosis and stroke-like episodes - MELAS)

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**UG HM – P201: Practical related to paper Fundamentals of Cell Biology & Physiology**

**30 hrs**

1	Study of prokaryotic cell structure and study of electron micrographs of all important cell organelles	<b>5 h</b>
2	Study of eukaryotic cell structure and study of electron micrographs of all important cell organelles	<b>5 h</b>
3	Micrometry- measurement of cell size taking different types of cell	<b>2 h</b>
4	Staining and observation of human cheek epithelial cells	<b>2 h</b>
5	Isolation and characterization of the following subcellular components using appropriate sample by differential centrifugation - nuclei (staining and counting), mitochondria (succinate dihydrogenase assay), Chloroplast (microscopic observation), lysosome (Acid phosphatase assay)	<b>6 h</b>
6	Methods of cell lysis and confirmation	<b>2 h</b>
7	Study of different stages of mitosis	<b>2 h</b>
8	Study of effects of colchicine on mitosis	<b>2 h</b>
9	Study of different stages of meiosis in <i>Tradescantia</i>	<b>2 h</b>
10	Study of polytene chromosomes ( <i>Drosophilla</i> /Chironomous larvae)	<b>2 h</b>

<b>Unit I</b>	<b>3 Hrs</b>	<b>Historical perspective</b> Origin of life with respect to abiotic production of biomolecules, cellular and chemical foundation of life- an overview
<b>Unit II</b>	<b>13 Hrs</b>	<b>Chemical foundation-(Overview)</b> a) Biomolecules as compounds of carbon with variety of functional groups b) Universal set of small molecules, macromolecules as the major constituents of cells: configuration and conformation with definitions and suitable example only, Types of stereoisomers and importance of stereoisomers in biology, types of bonds and their importance - electrovalent, covalent, ester, phosphodiester, thioester, peptide and glycosidic bonds
<b>Unit III</b>	<b>4 Hrs</b>	<b>Water</b> - properties of water, hydrogen bonding, structure ionization, interactions of biological molecules in water, osmosis, concept of pH and buffers, Buffering system in living cells
<b>Unit IV</b>	<b>20 Hrs</b>	<b>Carbohydrates</b> Definition, classification, biological role, structure, sugars and non-sugars, Monosaccharides- families of monosaccharides- aldoses, ketoses, trioses, tetraoses, pentoses and hexoses Definition, classification and brief account of monosaccharides (based on aldehyde and ketone groups), D and L configuration, mutarotation, epimers, anomers, chemical and physical, properties, glycosidic bond- properties and reaction of glucose and fructose-isomerism, oxidation and reduction, esterification and glycoside formation, osazone- structure of ribose, deoxyribose, glucose, galactose and fructose <b>Oligosaccharides and disaccharides-</b> concept of reducing non-reducing sugars, glycosides bonds, structure of lactose, sucrose, maltose, cellobiose, inversion of sugars <b>Polysaccharides-</b> its classification based on function- storage polysaccharides, homopolymers - starch and glycogens, heteropolymere - inuline, Structural polysaccharides- cellulose and chitin, peptidoglycan –functions of carbohydrates
<b>Unit V</b>	<b>20 Hrs</b>	<b>Lipids :</b> Blur's Classification, Storage and Structural lipids, Simple lipids (Triacylglycerol and waxes), Compound and complex lipids, phospholipids – phosphatidyl colin, ethanol amine, glycerolipids, sphingolipids, glycolipids, sterols, derived lipids, sphingomyline, cetebrosides, gangliosides, lipoproteins - LDL, VLDL, HDL; Lysosome Chylomicrones <b>Fatty acids –</b> nomenclature structure and properties (up to C18), Properties of lipids - Physical properties (state, colour, odour, melting point, solubility, specific gravity, geometric isomerism, emulsification and surface tension), Chemical properties (SAP value, Acid value, iodine number, rancidity),

		Functions of lipids
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**UG HM P202: Basics of Biochemistry – Biomolecules - I**

**30 hrs**

1	Biochemical calculations - preparation of solutions and buffers (pKa values) – w/v, v/v, %, ppm, ppb, mg/L, normality, molarity, molality	<b>3 h</b>
2	Study of colorimetry and preparation of standard graph and calculation of $\lambda_{\max}$ for given samples (Tyrosine/ purines/ pyrimidines), Verification of Beer-Lambert law by using Ammonium Copper compound, identification of purines from $\lambda_{\max}$	<b>3 h</b>
3	Isolation and identification of Starch from plant source	<b>2 h</b>
4	Saponification number - To find out saponification number of given lipid	<b>3 h</b>
5	Qualitative analysis for sugars and lipids	<b>2 h</b>
6	To estimate concentration of reducing sugar by DNSA method	<b>3 h</b>
7	To estimate concentration of Cholesterol in given sample (Iron reagent)	<b>2 h</b>
8	To separate and identify sugars by paper chromatography/ TLC	<b>2 h</b>
9	Detection of unknown carbohydrate from mixture (glucose, fructose, maltose, xylose, starch and sucrose)	<b>2 h</b>
10	To estimate reducing sugar from apple juice by Benedicts methods/Molish Test	<b>2 h</b>
11	Validation of glass pipettes and balance	<b>2 h</b>
12	Standardization of solution (0.25 N $K_2Cr_2O_7$ ) using 0.1 N ferrous ammonium sulphate and ferroin indicator	<b>2 h</b>
13	Determination of pH of different food samples by using pH paper/ universal pH standards	<b>2 h</b>

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**B. Sc. Part I Semester II**

**UG HM – T203: Basics of Biochemistry-Biomolecules - II**

<b>Unit I</b>	<b>20 Hrs</b>	<p><b>Proteins:</b></p> <p>i) Amino acids as building blocks of proteins, classifications of common amino acids (by R groups), uncommon amino acids and their functions, chemistry of amino acids, ionization of amino acid side chains, configuration, zwitterions, reactions of amino acids, titration of amino acids, isoelectric pH, reaction with Ninhydrin, Sanger reaction</p> <p>ii) Peptides and proteins: oligopeptides- structure and function of naturally occurring glutathione, insulin and synthetic aspartem Protein structure: importance of amino acid sequence; primary structures and concepts of N &amp; C terminal, peptide bond formation, characteristics of peptide bonds; Secondary structures: Ramchandran Plot, alpha helix and beta sheets, secondary repeats; tertiary and quaternary structure of protein (Haemoglobin), forces holding the polypeptides together - hydrogen bonds, Vanderwaals forces, covalent, ionic bonds and salt linkages; Protein denaturation and renaturation; Classification of protein shape, structural, transport, chromosomal, phospho and glyco proteins and the biological role of proteins.</p>
<b>Unit II</b>	<b>10 Hrs</b>	<p><b>Nucleic acids:</b></p> <p>Occurrence, purines, pyrimidines, Pentoses (Ribose and Deoxyribose) phosphates, AMP and cAMP, ADP and ATP, TDP and TTP, GDP and GTP, NDA, NADP, FMN and FAD; Polynucleotides, covalent structure of DNA (different forms of DNA) and RNA (mRNA, tRNA, rRNA and SnRNA); Forces stabilizing nucleic acid structures, N-β glyosidic bonds, Phosphodiester bonds,</p> <p>Properties of nucleic acids, denaturation and renaturation, Watson and Crick's model of DNA structure, ribozyme, Biological role of nucleic acids</p>
<b>Unit III</b>	<b>10 Hrs</b>	<p><b>Vitamins:</b></p> <p>Occurrence and sources, rich sources of different Vitamins, classification, structure &amp; biochemical functions of water soluble vitamins; Role as coenzymes: Thiamine, Riboflavin, Niacin, Pyridoxine, Pantothenic acid, Coenzyme A, Lypoic acid, Folic acid and B12; functions and deficiency symptoms</p>
<b>Unit IV</b>	<b>5 Hrs</b>	<p><b>Minerals:</b></p> <p>Role of Na, K, Mg, Fe, Zn, Co, Ca, P and I in physiology, general electronic configuration and their shape and significance in metalloenzymes</p>
<b>Unit V</b>	<b>10 Hrs</b>	<p><b>Enzymes :</b></p> <p>Definition, structure and concept of Apoenzyme, Coenzyme, Cofactor Prosthetic group, Active site, Types of enzyme, Extracellular and intracellular, Constitutive and inducible, general overviews of enzyme-substrate reaction, mechanism of enzyme action, factors affecting enzyme</p>

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		reactions
<b>Unit VI</b>	<b>5 Hrs</b>	<b>Plant Pigments and Dyes:</b> Chlorophyll, Xanthophylls, Flavonoids, Carotenes, etc.

**UG HM – P203 Practical: Basics of Biochemistry-Biomolecules - II**

**30 hrs**

1	Estimation of concentration of protein by Biuret method and Lowry method (Albumin)	<b>2 h</b>
2	Study of melting temperature of nucleic acid- to determine T <sub>m</sub> of DNA and mole percent G+C content	<b>2 h</b>
3	To separate amino acid by TLC	<b>2 h</b>
4	To study amylase enzyme assay- and to study effects of pH, temperature, concentration of enzyme, activators and inhibitors	<b>2 h</b>
5	General tests for amino acids and detection of unknown amino acid from mixture (Arginine, cysteine, methionine, Tyrosine, histidine, proline and tryptophan)	<b>2 h</b>
6	Isolation and characterization casein from milk by Isoelectric pH method	<b>2 h</b>
7	Estimation of DNA by DPA and RNA by Orcinol methods	<b>2 h</b>
8	Preparation of titration curve of acidic, basic and neutral amino acids	<b>2 h</b>
9	Quantitative estimation of ascorbic acid	<b>2 h</b>
10	Bioassay of Vitamin B12	<b>2 h</b>
11	Separation of pigment and dyes by adsorption and ion exchange chromatography	<b>2 h</b>
12	Extraction of genomic DNA from onion/yeast/ rat liver/ bacteria and confirmation with DPA and agarose gel electrophoresis	<b>2 h</b>
13	Study of karyotype analysis (karyotyping)	<b>2 h</b>
14	Detection of significant industrial enzymes (amylase, protease, lipase, invertase, phosphatase and cellulase)	<b>2 h</b>
15	Enzymatic preparation of biomolecules - Dextrin- production of maltodextrin by using $\beta$ amylase Glucose- Productive of glucose by bacterial $\alpha$ - amylase and amyloglucosidase Production of invert sugar by invertase Peptide preparation of proteolysis by using papain Softening of Chhole/Rajma/ Idli by using papain	<b>2 h</b>

**UG HM T204: Microbial Nutrition, Growth and Bioenergetics**

<b>Unit I</b>	<b>20 Hrs</b>	<p><b>Chemical composition of microbial cell</b></p> <ul style="list-style-type: none"> <li>• Nutritional requirements: Carbon, Oxygen and Hydrogen, Nitrogen, Sulphur and Phosphorous, Minerals, growth factors and energy source -auxotroph, prototroph and fastidious microorganisms</li> <li>• Classification/categories of microorganisms</li> <li>• Microbial Nutrition, Cultivation and Isolation and Preservation</li> <li>• Design and preparation of culture media, Types of culture media - liquid and solid media, synthetic/ chemically defined media, semisynthetic complex non synthetic media, anaerobic growth media, selective and deferential media, indicator media, transport media; enrichment, isolation and pure culture techniques for microorganisms</li> <li>• Methods of purification of microorganisms - streak plate, spread plate, pour plate techniques, single cell isolation technique</li> <li>• Preservation of microbial cultures – slants, slants + mineral oil overlay, butt method (stabs), cryopreservation, freeze drying method (ampoules)</li> </ul>
<b>Unit II</b>	<b>5 Hrs</b>	<p><b>Overviews of culture collection centres and their role:</b> Requirements and guidelines of National Biodiversity Authority (NBA) for culture collection centres</p>
<b>Unit III</b>	<b>20 Hrs</b>	<p><b>Microbial growth:</b> Inoculation techniques and study of growth - Inoculation of liquid medium (broth), Solid media (slants, butts and plates), Study of colony characteristics of pigment and pigment non producing bacteria, Study of motility- hanging drop preparation and sloppy agar method, Kinetics of bacterial growth (exponential growth model), phases of growth, Growth curve - generation time, continuous (exponential), Chemostat, diauxic and synchronous growth Measurement of microbial growth methods of enumeration</p> <p>a) Microscopic methods (Direct microscopic count, haemocytometry method), counting cells using improved Neubauer-Petroft-Hosser's chamber</p> <p>b) Plate count (serial dilution technique) - total viable count/SPC/Breed's smear count, membrane filtration technique</p> <p>c) Turbidometric method- Nephelometry/ Electronic counter method (Coulter counter) Tetrazolium chloride method</p> <p>d) Brown's opacity tube method/MBRT and Resazurine estimation of biomass (dry mass packed cell volume)</p> <p>e) Chemical methods- Cell carbon and nitrogen estimation Determination of optimum growth conditions – pH, temperature, solute concentration (salt, sugars), heavy methods and incubation period</p>
<b>Unit IV</b>	<b>5 Hrs</b>	<p><b>Microbial growth in natural environments:</b> Soil, Water, Food, Animal and Plant body, Microbial Parasites) Methods for cultivation of photosynthetic, extremophilic and chemolithotropic (chemoorganotrops) bacteria, anaerobic bacteria, algae, fungi (yeast and molds), protozoa, actinomycetes and viruses</p>
<b>Unit V</b>	<b>10 Hrs</b>	<p><b>Bioenergetics:</b> Principle of bioenergetics, Role of ATP in metabolism, reducing power and its</p>



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		significance in metabolism, generation of ATP through substrate level phosphorylation, components of electrons transport chain (ETC)- Flavoproteins (FMN, FAD), Quinines (Ubiquinones, Menaquinons), Iron sulphur proteins, cytochromes - generation of ATP through ETC
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<b>Sr. No.</b>	<b>Practical</b>	<b>Hours</b>
<b>1</b>	Introduction & use of common laboratory glasswares / labwares – testtubes, culture tubes, suspension tube, screw capped tubes, Petriplate, Pipettes (Mohr & serological) Micropipettes,, Pasteur pipettes, Erleyer meyar flasks, Volumetric flasks, Glass spreaders, Durham’s tubes, Cragie’s tube & inoculating needle (wire loops, Stab needles)	<b>2 h</b>
<b>2</b>	Learning basic techniques in Microbiology – Wrapping of glasswares, cotton plugging, cleaning & washing of glassware, biological waste disposal	<b>1 h</b>
<b>3</b>	Preparation of simple laboratory media - nutrient agar, broth, Mac-Conkey’s agar, Manitol salt agar, Peptone water, Sabouraud’s agar & their sterilization, checking of sterilization efficacy of autoclave using biological indicator ( <i>Bacillus stearothermophilus</i> )	<b>2 h</b>
<b>4</b>	Study of motility by hanging drop method and study of swarming phenomenon on sloppy agar medium	<b>1 h</b>
<b>5</b>	Preparation of Winogradsky’s column & observation of different types of microorganisms using bright field microscope	<b>1 h</b>
<b>6</b>	Pure culture techniques – Streak, spread, pour plate methods & study of colony characteristics	<b>2 h</b>
<b>7</b>	Isolation, colony characteristics, gram staining, motility of following bacteria – <i>E. coli</i> , <i>Bacillus spp.</i> , <i>Staphylococcus spp.</i> , <i>Micrococcus spp.</i> , pigment & pigment non producing microorganisms	<b>1 h</b>
<b>8</b>	Wet mount and slide preparation for algae, fungi & protozoa using sample sources for Amoeba spp., Paramecium spp., Nostoc, Chlorella, Aspergillus, Mucor & Penicillium, Fusarium, <i>Rhizopus spp.</i>	<b>2 h</b>
<b>9</b>	Inoculation techniques & study of growth in liquid broth media, solid media, slants, butts & plates, coverslip & slide culture techniques for actinomycetes	<b>1 h</b>
<b>10</b>	Effect of environmental factors on growth of bacteria ( <i>E. coli</i> , <i>Staphylococcus aureus</i> ) - pH, temperature, salt concentration, heavy metals (oligodynamic action)	<b>1 h</b>
<b>11</b>	Study of normal flora of skin – observing & cultivating different morphoforms of microorganisms from skin & effect of washing of skin with soap & disinfectant on microflora	<b>2 h</b>
<b>12</b>	Preservation of culture on slants, in soil & on grain surfaces, butts, vials/ampoules/lyophils & revival of these cultures & lyophils	<b>2 h</b>
<b>13</b>	Enrichment, isolation & morphological studies of – Chemoautotrophs, Chemoorganotrops , Photoautotrophs, Photoorganotrops (one member each)	<b>2 h</b>
<b>14</b>	Study of growth curve, continuous growth / diauxic / synchronous growth	<b>2 h</b>
<b>15</b>	Measurement of bacteria by Direct Microscopic Count (DMC), Slide / Neubauer’s chamber, direct plating (SPC) , Indirect – Nephelometry / Brown’s opacity tube / MBRT	<b>2 h</b>
<b>16</b>	Estimation of ATP generation	<b>2 h</b>
<b>17</b>	Cultivation of anaerobic bacteria from natural sources	<b>2 h</b>

**B. Sc. Part I Semester II**  
**UG HM -T205 Advanced Chemistry, Physics & Biophysics for Biologists**

Unit	Hours	Topics
Unit I	5	<b>Chemistry of transition &amp; non transition elements</b> ❖ Transition elements – General properties (d & f block elements), electronic configuration, oxidation state, magnetic movement & complexes of 3d & lanthanide elements ❖ Non – transition elements – General properties (s & p block elements); synthesis, properties & structure of halides & oxides of Carbon, silicon & Nobel gas compounds
Unit II	4	<b>Colloidal state</b> Colloidal system, classification & size range of colloids, preparation & purification of colloidal solutions, general properties of colloidal system, some properties of hydrophobic colloidal system (electrical & electrokinetics), Surfactants, emulsions, Gels, importance & applications of colloids
Unit III	7	<b>Electrochemistry</b> – Introduction, electrochemical cell, cell constant, half cell & potential reaction, reduction potential, transport number, conductance, Kohlrausch law, electrochemical series, thermodynamics, potential function from cell, potential measurement & it's applications, Emf, Nernst's equation, Galvanic cells, Liquid – junction potential, Huckel theory, over voltage / over potential Bioelectricity – Introduction, electricity observed in living system – examples, origin of bioelectricity, resting potential & action potential, conduction velocity, pace maker, ECG, EEG, EMG, EOG
Unit IV	4	<b>Name reactions</b> – Introduction, Mannich reaction, Hoffmann reaction, Diels – Alder reaction, Perken's reaction, Meerwein – Ponndorf – Verley (MPV) reduction
Unit V	3	<b>Elasticity</b> – Basic concept of stress & strain in solids, Hook's law, stress, strain curve, properties of fluids
Unit VI	3	<b>Thermometry</b> – Principles of thermometry, concept of temperature & it's measurement, Thermal energy, Platinum resistant thermometer, thermocouple, thermistors as thermometer
Unit VII	6	<b>Conventional &amp; non- conventional energy sources &amp; devices</b> – Introduction various types of conventional & non-conventional energy sources – Solar energy, direct use of solar energy – Silicon solar cells, principle of conversion of solar energy in to electricity & construction of solar cell (spectral distribution), efficacy, fill factor
Unit VIII	4	<b>Ideal &amp; real gases</b> Ideal gas – Kinetic model, gas equation, kinetic interpretation of temperature, degree of freedom, equipartition of energy, real gas – deviation of behaviour of real gases from the ideal gases, critical constants of a gas ( $P_c$ , $V_c$ & $T_c$ ), Vanderwaal's equation, liquification of gases.
Unit IX	3	<b>Current electricity</b> – Introduction, active & passive components, A. C., L-R, R-C, C-R circuits, half wave rectifier, full wave rectifier, bridge rectifier & transformers

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<b>Unit X</b>	<b>4</b>	<b>Semiconductors</b> Introduction, definition & examples of conductor, semiconductor, insulator, intrinsic & extrinsic semiconductors, types of semiconductor diodes, Pn junction diode, Zener diode, Transistors – p-n-p & n-p-n transistors, common emitters & best circuits, light emitters diode (LED) and segment display, photodiode, optocoupler
<b>Unit XI</b>	<b>5</b>	<b>Optics</b> Introduction, interference, in parallel test thin films, wedge – shaped thin films, Newton’s rings, Polarization of light & concept of optical activity, diffraction - types, diffraction – grating, experimental, determination of wavelength by diffraction grating, Lasers – properties, Lasers action, (energy level diagram), Concept of population inversion, optical pumping & Einstein’s equation, Nicol’s prism properties, Rubby laser
<b>Unit XII</b>	<b>3</b>	<b>Introduction to digital electronics</b> <ul style="list-style-type: none"><li>• Number system &amp; logic gates</li><li>• Small signal voltage amplifiers, number systems – decimal, binary, BCD, Basic logic gate, bit groupings, CoR, NoR, AND, NAND, NoT, DeMorgan’s theorem, Half adder &amp; full adder</li></ul>
<b>Unit XIII</b>	<b>3</b>	<b>Magnetism</b> Magnetic field, maghetism of earth, para, dia, ferro, nuclear & biomagnetism
<b>Unit XIV</b>	<b>4</b>	<b>Overview of green chemistry &amp; synthesis</b> – Microwave assisted synthesis of organic compounds, retrosynthesis

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**UG HM -P205 Practical: Advanced Chemistry, Physics & Biophysics for Biologists**  
**(30 hrs)**

<b>Sr. No.</b>	<b>Practical</b>	<b>Hours</b>
<b>1</b>	Determination and adjustment of pH of solutions	<b>2 h</b>
<b>2</b>	Preparation of different buffer solutions	<b>2 h</b>
<b>3</b>	Determination of heat of solution of Benzoic acid / Salicylic acid by solubility measurements	<b>2 h</b>
<b>4</b>	Estimation of acetone by idometric titration method	<b>2 h</b>
<b>5</b>	Determination of conductivity of solutions	<b>3 h</b>
<b>6</b>	Determination of Optical activity by polarimeter	<b>3 h</b>
<b>7</b>	Study of depression in freezing point	<b>3 h</b>
<b>8</b>	Determination of dissociation constant of weak acid Study of substituent on dissociation constant of weak acid	<b>3 h</b>
<b>9</b>	Inorganic estimation of amount of magnesium from talcum powder by complexometric titration	<b>2 h</b>
<b>10</b>	Study of principle, working & construction of pH meter & conductivity meter	<b>2 h</b>
<b>11</b>	Demonstration of principle, working & construction of Refractometer, Laminar Air Flow	<b>4 h</b>
<b>12</b>	Purification of any two organic compound by recrystallization selecting suitable solvent	<b>2 h</b>

**B. Sc. Part I Semester II**  
**UG HM-T206 Applied Plant and Animal Sciences**

Unit	Hours	Topics
<b>Unit I</b>	<b>10</b>	<b>Plant water relationship and its importance</b> Definition, significance and mechanism: i. Permeability; ii. Diffusion & imbibitions; iii. Osmosis & its types Relation between osmotic pressure (OP), turgor pressure (TP) and wall pressure (WP), Diffusion Pressure Deficit - DPD (Suction pressure) Absorption and Transport of water: Introduction and mechanism of Ascent of sap - transpiration and guttation, Translocation of mineral elements (Capillarity, Imbibition, Atmospheric pressure and Cohesion-tension)
<b>Unit II</b>	<b>4</b>	<b>Plant Metabolism:</b> Photosynthesis: - Photosynthesis pigments, concept of two photo systems, photophosphorylation, Calvin cycle, CAM (Crassulacean Acid Metabolism) plants, photorespiration, compensation point. Respiration: Mechanism - Glycolysis, Krebs's cycle and ETS Nitrogen metabolism- inorganic & molecular nitrogen fixation
<b>Unit III</b>	<b>4</b>	<b>Growth and development of plants :</b> Essential nutrients for Plant growth and their role Plant growth regulators Introduction to physiology of flowering: a) Photoperiodism b) Vernalisation Economic importance of plants: Cereals, Pulses, Oil seeds, Fiber plants, Medicinal Plants, Timber yielding, Beverages with examples
<b>Unit IV</b>	<b>10</b>	<b>Animal Physiology</b> Digestion: Structure and function of digestive glands; Digestion and absorption of carbohydrates, fats and proteins Respiratory: Physiology, External and internal Respiration, Transport of oxygen and carbon dioxide in blood, Factors affecting transport of gases. Functioning of Excitable Tissue (Nerve and Muscle) - Structure of neuron, Propagation of nerve impulse (myelinated and nonmyelinated nerve fibre); Structure of skeletal muscle, Mechanism of muscle contraction (sliding filament theory), Neuromuscular junction Endocrine and Reproductive Physiology - Structure and function of endocrine glands (pituitary, thyroid, parathyroid, pancreas, adrenal, ovaries, and testes), Brief account of spermatogenesis and oogenesis
<b>Unit V</b>	<b>3</b>	<b>Parasitology</b> Introduction to Host-parasite Relationship - Host, Definitive host, Intermediate host, Parasitism, Symbiosis, Commensalism Parasitic Protozoa: Life history and pathogenicity of <i>Plasmodium vivax</i> Parasitic Helminthes: Life history and pathogenicity of <i>Fasciola hepatica</i> , <i>Taenia solium</i>
<b>Unit VI</b>	<b>2</b>	<b>Economic Zoology</b> Vermiculture; Aquaculture; Sericulture and Apiculture
<b>Unit VII</b>	<b>4</b>	<b>Conventional &amp; non- conventional energy sources &amp; devices</b> Introduction - various types of conventional & non-conventional energy sources – Solar energy, direct use of solar energy – Silicon solar cells, principle of conversion of solar energy in to electricity & construction of solar cell (spectral distribution), efficacy, fill factor
<b>Unit VIII</b>	<b>4</b>	<b>Ideal &amp; real gases</b>

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		Ideal gas – Kinetic model, gas equation, kinetic interpretation of temperature, degree of freedom, equipartition of energy, real gas – deviation of behaviour of real gases from the ideal gases, critical constants of a gas ( $P_c$ , $V_c$ & $T_c$ ), Vanderwaal's equation, liquification of gases.
<b>Unit IX</b>	3	Current electricity – Introduction, active & passive components, A. C., L-R, R-C, C-R circuits, half wave rectifier, full wave rectifier, bridge rectifier & transformers
<b>Unit X</b>	4	<b>Semiconductors</b> Introduction, definition & examples of conductor, semiconductor, insulator, intrinsic & extrinsic semiconductors, types of semiconductor diodes, Pn junction diode, Zener diode, Transistors – p-n-p & n-p-n transistors, common emitters & best circuits, light emitters diode (LED) and segment display, photodiode, optocoupler
<b>Unit XI</b>	5	<b>Optics</b> Introduction, interference, in parallel test thin films, wedge – shaped thin films, Newton's rings, Polarization of light & concept of optical activity, diffraction - types, diffraction – grating, experimental, determination of wavelength by diffraction grating, Lasers – properties, Lasers action, (energy level diagram), Concept of population inversion, optical pumping & Einstein's equation, Nicol's prism properties, Rubby laser
<b>Unit XII</b>	3	<b>Introduction to digital electronics</b> <ul style="list-style-type: none"> <li>• Number system &amp; logic gates</li> <li>• Small signal voltage amplifiers, number systems – decimal, binary, BCD, Basic logic gate, bit groupings, CoR, NoR, AND, NAND, NoT, DeMorgon's theorem, Half adder &amp; full adder</li> </ul>
<b>Unit XIII</b>	3	<b>Magnetism</b> Magnetic field, maghetism of earth, para, dia, ferro, nuclear & biomagnetism
<b>Unit XIV</b>	4	Overview of green chemistry & synthesis – Microwave assisted synthesis of organic compounds, retrosynthesis

<b>Sr. No.</b>	<b>Practical</b>	<b>Hours</b>
<b>1</b>	Study the process of Osmosis and Turgor pressure and determination of Diffusion Pressure Deficit	<b>3 h</b>
<b>2</b>	Determination of rate of respiration	<b>3 h</b>
<b>3</b>	Estimation of chlorophyll content in photosynthesizing and non photosynthesizing leaf	<b>3 h</b>
<b>4</b>	Effect of plant growth regulators on germination of seeds	<b>4 h</b>
<b>5</b>	Studies on economically important plants: Students should prepare herbarium specimens with their uses	<b>3 h</b>
<b>6</b>	Study and dissection of Honey Bee , Mounting of Mouth parts, pollen basket, Antenna Cleaner, Sting Apparatus , legs and wings	<b>3 h</b>
<b>7</b>	Study of Plasmodium spp.	<b>3 h</b>
<b>8</b>	Study of Fasciola sp.	<b>3 h</b>
<b>9</b>	Enumeration of red blood cells using haemocytometer.	<b>3 h</b>
<b>10</b>	Collection, Classification and preservation of Insects - Drosophila	<b>2 h</b>



**UG HM T207: Ecology, Ecosystem & Geosciences**

Unit	Topics	Hours
<b>Unit I</b>	<p>Fundamentals of ecology</p> <ul style="list-style-type: none"> <li>• Environments: definition, components –                             <ol style="list-style-type: none"> <li>a) Atmosphere - origin, composition, structure, variables</li> <li>b) Hydrosphere – Characteristics, hydrological cycle, El Nino, La Nina</li> <li>c) Lithosphere – Formation, zonal structure, soil studies – origin, profile, properties, classification</li> <li>d) Biosphere – Characteristics &amp; inter-relationships</li> </ol> </li> <li>• Ecological spectrum &amp; hierarchy, levels of organization, autecology, synecology, population, community, biomes &amp; ecosystem ecology.</li> </ul>	10
<b>Unit II</b>	<p>Ecosystem structure &amp; function –                      Concept of ecosystem, types of ecosystem structure – biotic &amp; abiotic components, Macro &amp; micro ecosystem                      Function – a) Food chain – Grazing, detritus                      b) Food web &amp; ecosystem stability, Trophic levels                      c) Ecological energetics – Energy input / Energy flow (Single channel &amp; Y shaped models)                      d) Productivity of ecosystem – Primary production (GPP &amp; NPP), Secondary production, Standing crop (biomass)                      e) Ecological pyramids – Number, biomass &amp; energy.</p>	15
<b>Unit III</b>	<p>Biogeochemical cycles –</p> <ul style="list-style-type: none"> <li>• Nutrient cycling –                             <ol style="list-style-type: none"> <li>a) Gaseous cycle - Hydrological, Carbon, nitrogen, Oxygen</li> <li>b) Sedimentary cycle – Phosphorus, sulphur, Calcium &amp; Magnesium</li> </ol> </li> <li>• Ecosystem nutrient cycling modes – Intra – system cycling &amp; extra system transfer – Nutrient inputs, biotic accumulation of nutrients, nutrient outputs</li> </ul>	10
<b>Unit IV</b>	<ul style="list-style-type: none"> <li>• Population ecology – Introduction, basic concept, population characteristics – size &amp; density, dispersion (random, aggregate &amp; uniform) nativity (potential &amp; realized), fecundity, mortality (potential &amp; realized), survival curve, age &amp; sex structure, life table &amp; viability analysis, concept of carrying capacity</li> <li>• Population growth – a) Growth curves exponential &amp; logistic                              b) Population fluctuation                              c) Biotic potential &amp; environmental resistance</li> </ul>	10
<b>Unit V</b>	<ul style="list-style-type: none"> <li>• Community ecology – Characteristics of commonly – Species diversity, growth forms &amp; structure, Dominance, succession, trophic structure, ecological Niche, ecotone &amp; edge effect</li> <li>• Characters in community structure – Analytic (Qualitative &amp; Quantitative) &amp; synthetic</li> <li>• Inter – specific &amp; intra – specific relationships</li> <li>• Concept of succession, causes of succession, basic types – primary, secondary, autogenic, allogenic etc.</li> <li>• Mechanism of succession – Nudation, invasion, competition, Co-action &amp; reaction, stabilisation (climax), models &amp; succession – Hydrosere &amp; lithosere</li> </ul>	10

<b>Unit VI</b>	Threats to the environment & ecosystem	5
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**Practical P207: Ecology, Ecosystem & Geosciences**

**30 hrs**

Sr. No.	Practical	Hours
<b>1</b>	Study of ecosystem (Aquatic, forest, river etc.)	4 h
<b>2</b>	Community sampling by quadrat methods for plants – Percentage of frequency, density, abundance, frequency class diagram & comparison with Raunkiaer’s frequency chart, Simpson’s index & dominance, Shannon diversity index	6 h
<b>3</b>	Measurement of primary productivity of grassland by harvest method	4 h
<b>4</b>	Determination of frequency, abundance (Line) & density (Belt) of species across terrestrial – aquatic transitional zones	5 h
<b>5</b>	Case studies on ecological succession	3 h
<b>6</b>	Study of natural resources Forest / Mineral / Food / Water / Land	2 h
<b>7</b>	Study of ecological pyramids	2 h
<b>8</b>	Study of different food chains	2 h
<b>9</b>	Field visits	2 h

**B. Sc. Part I Semester II**

**UG HM- T208 Basics of Environmental Pollution and Applied Microbiology & Food and Dairy**

Unit	Topics	Hours
<b>Unit I</b>	<b>Environmental Pollution &amp; control:</b> Introduction, definitions, sources & types of pollution	<b>4</b>
<b>Unit II</b>	<p><b>Water pollution &amp; microbiology:</b> Sources &amp; classification of water pollution, different types of aquatic environments, water pollution parameters &amp; their biological significance:</p> <ul style="list-style-type: none"> <li>▪ Physical – Colour, odour, temperature, turbidity &amp; density</li> <li>▪ Chemical – Solids (suspended, total &amp; dissolved, volatile), Hardness, acidity, alkalinity, pH, DO, ions (Fe, Cu, Mn, Na, K, Ca, N, P, F, Cl)</li> <li>▪ Pollutants – Chemicals, pesticides &amp; detergents</li> <li>▪ Biological coliforms (faecal, streptococci), Organic matter (BOD, COD) &amp; their significance as pollution indicators</li> <li>▪ Thermal pollutants – Waste heat &amp; it's uses, cooling ponds &amp; towers, effect of thermal pollution on light &amp; atmosphere</li> <li>▪ Normal flora of water, sources of microorganisms in water, faecal pollution, most prominent waterborne pathogens, indicators of faecal pollution</li> <li>▪ Water quality assays – routine bacteriological examination of water (SPC) test for coliforms</li> <li>▪ Qualitative (preventive, confirmed &amp; completed tests), IMViC test, Eijkman test, Quantitative – MPN, Membrane filter technique</li> <li>▪ Treatment &amp; purification (primary-physical, secondary-biological &amp; tertiary-chemical) of municipal drinking water supply</li> <li>▪ Eutrophication</li> <li>▪ Groundwater &amp; marine pollution.</li> </ul>	<b>20</b>
<b>Unit III</b>	<p><b>Air pollution &amp; aeromicrobiology</b></p> <p>Compassion of air, types &amp; classification of air pollutants, gaseous inorganic air pollutants – NO<sub>x</sub>, SO<sub>x</sub>, CO, CO<sub>2</sub>, H<sub>2</sub>S, NH<sub>3</sub>, O<sub>3</sub>, CFC.</p> <ul style="list-style-type: none"> <li>▪ Organic air pollutants – aliphatic &amp; aromatic organic compounds, particulate matters, types &amp; effects</li> <li>▪ microbial pollutants – number &amp; types of microorganisms in air, sources, infectious dust –droplets &amp; droplets nuclei, microbiological examination of air – air samplers &amp; samplings methods – solid impaction (sieve device) &amp; liquid impingement – (bead bubbler device).</li> <li>▪ Acid rain, photochemical SMOGs, London &amp; LA SMOGs (mechanisms of formation) decrease of ozone layer (role of CFC's &amp; control).</li> <li>▪ Green house effects, instrumental analysis of SO<sub>x</sub>, NO<sub>x</sub>,</li> </ul>	<b>15</b>

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	<p>economic impact of air pollutant</p> <ul style="list-style-type: none"> <li>▪ Effect of air pollution of human, plants, animals &amp; atmospheric health</li> </ul>	
<b>Unit IV</b>	<p>Soil pollution &amp; Microbiology:            Definition, sources, role of pesticides in soil pollution.            Soil types, types of microbes found in soil, role of microorganisms in soil fertility, soil pollution control measures.</p>	<b>6</b>
<b>Unit V</b>	Noise pollution – Sources & types of noise, sonic boom, measurements of noise effects & control measures	4
<b>Unit VI</b>	Radiation pollution – Introduction, atomic radiations, effect of radiation, radioactive waste & disposal, radiation protection	4
<b>Unit VII</b>	<p>Environmental toxicology – Definition, classification &amp; concept.            Pesticide toxicity (organic &amp; inorganic), mode of action of toxicants of metals – arsenic, mercury, cadmium, lead, Nickel, Asbestos, chromium, organo phosphate, carbamates, etc., mutagens &amp; carcinogens, Cyanide, Peroxy Acetyl Nitrate (PAN), dioxins.            Bioconcentration, bioaccumulation, Biomagnification, potentiation &amp; Synergism            Control of toxic effect, biotransformation &amp; excretion</p>	4
<b>Unit VIII</b>	Energy – Renewable & Non-renewable energy sources, fossil fuels, CNG, Crude oil, Coal, fractional distillations of crude oil, bioethanol from sugary & starchy crops, petrocrops – rubber, Biodiesel (production, advantages & limitations)	3

**Practical P208 Basics of Environmental Pollution and Applied Microbiology & Food and Dairy**

**30 hrs**

<b>Sr. No.</b>	<b>Practical</b>	<b>Hours</b>
<b>1</b>	Determination of temporary & permanent hardness of water	<b>2</b>
<b>2</b>	Estimation of COD & DO, BOD of polluted water samples	<b>2</b>
<b>3</b>	Determination of solid content of polluted water samples (SS, TS, DS, VS)	<b>2</b>
<b>4</b>	Routine bacteriological analysis of water – preventive, confirmed & completed test, MPN, Eijckmen's Test	<b>2</b>
<b>5</b>	Bacteriological analysis of water - IMViC test	<b>2</b>
<b>6</b>	Study of degradation of pesticides using microorganisms	<b>2</b>
<b>7</b>	Enumeration of microorganisms from air by solid impaction & liquid impingement techniques	<b>2</b>
<b>8</b>	Study of effect of pesticides on azotobacter population by viable count method	<b>2</b>
<b>9</b>	Study of effect of heavy metals on growth of microorganisms	<b>2</b>
<b>10</b>	Estimation of noise by dB meter ( $L_{eq}$ )	<b>2</b>
<b>11</b>	Determination of nitrate & phosphate content in polluted water	<b>2</b>
<b>12</b>	Determination of PM concentration using High Volume Air Sampler (HVS)	<b>2</b>
<b>13</b>	Determination of organic matter and carbon from given soil sample	<b>2</b>
<b>14</b>	Determination of chlorine demand for the potable water	<b>2</b>
<b>15</b>	Detection of radioactive material in fruits & vegetables	<b>2</b>

<b>Unit</b>	<b>Topics</b>	<b>Hours</b>
<b>Unit I</b>	<b>Communication as part of science:</b> Language – a means of Communication; Communication – Meaning of Communication, Definitions; Principles of communications; Communication – Situation for and need of communication, Importance of communication Features, objectives and functions of communication, Communication cycle, Elements of Communication, Communication process, stages in Communication process	<b>5</b>
<b>Unit II</b>	<b>Types of Communications:</b> Formal – Informal, Verbal – Nonverbal, Vertical – Horizontal Diagonal	<b>2</b>
<b>Unit III</b>	<b>Principles of effective communication</b> Definitions of effective communication; Communication barriers and ways to overcome them; Developing effective messages – Knowledge about the audience, purpose of communication, structure of message, selecting the proper channel, avoiding barriers in communication, facilitating feedback.	<b>4</b>
<b>Unit IV</b>	<b>Non -Verbal Communication</b> Non – verbal codes: Body Language, chronemics and Artifacts	<b>1</b>
<b>Unit V</b>	<b>Illustrating with visuals:</b> Photographs, tables, graphs, flow charts, figures, maps, picture diagrams, pie diagrams, family tree.	<b>1</b>
<b>Unit VI</b>	<b>Formal written skills</b> i. Report writing: Seminar report, Conference report, Progress report, Investigative report, Accident report, Fall/rise in the Production, Joining report ii. Applications: Job Application with resume (C.V.), Sick leave application, Application for getting particular information (eg. prospectus / prescribed admission / scholarship form). iii. Business correspondence: Enquiry letter, Order letter, Complaint letter, Adjustment Letter iv. Office drafting: Circular, Notice, Memo, Defining and Describing object and Giving Instructions	<b>2</b>

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**UG HM- T210 Personality Development**

Unit	Topics	Hours
<b>Unit I</b>	<b>Planning and Goal setting:</b> Five skills needed to achieve carrier goals: Human perceptions, Understanding people, types of soft skills, Types of soft skills, Need for achievement and Spiritual Intelligence, Developing potential and self actualization	<b>5</b>
<b>Unit II</b>	<b>Conflicts and stress:</b> Types of conflicts, conflict resolution skills, Types of stress, causes of stress, effects of stress and regulating the stress; Habits – Good and bad habits, Forming Habits of success, breaking bad habits.	<b>3</b>
<b>Unit III</b>	<b>Communication skills</b> Communication cycle advanced and essentials, Basic telephonic skills. Communication barriers- Interpersonal transactions, miscommunication Technology and Communication - Email- Principle, Netiquettes, E-mail etiquettes	<b>4</b>
<b>Unit IV</b>	<b>Presentation skills:</b> Overcoming fear, Becoming a professional, the role of body language, effective reading and using visuals.	<b>3</b>

**UG HM – T211VAC: Introduction to Research Methodology – II**

<b>Unit- I</b>	<p><b>Scientific Writing-</b></p> <ol style="list-style-type: none"> <li>1) Language as means of communication – English language</li> <li>2) Scientific writing verses unscientific writing- Scientific writing in English language</li> <li>3) Good English and grammar in scientific writing - Basic grammar, Tenses, Voices, Prepositions and Conjunctions, conditional sentences, count and non count nouns, concord and punctuations, use and misuse of words, jargons and avoiding jargons, use of abbreviations, accepted abbreviations and symbols, common error in the style and in spellings.</li> <li>4) Scientific methods – Concept, hypothesis, theory, law, design of experiment, inductive &amp; deductive reasoning.</li> <li>5) General structure of scientific reports (types of scientific documents) – Journal articles, books, posters, conference, papers, thesis, review papers, books reviews, project &amp; conference reports.</li> <li>6) Writing a scientific papers – IMRAD/IRDAM acronym/ system, literature search, title, listening of authors &amp; addresses, abstract, key words, introduction, material –method, result &amp; discussion, summary &amp; conclusion, references, stating the acknowledgement, tables/graphs/diagrams &amp; illustrations</li> <li>7) Structure of project – Title, author &amp; their institution, abstract/ summary, certificates (students undertaking, guide certificate,</li> </ol>	<b>15 hrs</b>
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	plagiarism checker certificate, ethical clearance), acknowledgements, list of content, abbreviations, introduction, literature survey, aim & objectives, material & methods, results & discussion, conclusion/recommendation, bibliography, annexure (list of chemicals, glasswares, reagents, media used with composition, paper publication etc.).	
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**UG HM – P211VAC: Practical course Research methodology**

**30 hrs.**

1	Writing suitable title of research papers, search of instructions to authors from website of scientific journal (its analysis and comparison)	02 hrs.
2	Writing abstract for research paper	02 hrs.
3	Writing summary and conclusion for given scientific paper	
4	Writing a bibliography for given research paper	02 hrs.
5	Preparation of research paper for publication (may be on their research project)	08hrs.
6	Prepare a plagiarized and non plagiarized document (use of plagiarism checker)	03 hrs.



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